

October 25, 2022

Bernadette Juarez  
 APHIS Deputy Administrator  
 Biotechnology Regulatory Services

**RECEIVED**

By kldiggs for BRS Document Control Officer at 9:23 am, Oct 26, 2023

Ms. Juarez,

With this letter we respectfully request a Regulatory Status Review from USDA-APHIS' Biotechnology Regulatory Services (BRS) for the following three Plant Trait Mechanisms of Action: *Thlaspi arvense* L. (pennycress; field pennycress) CRISPR/Cas9 generated mutant lines, featuring disruption of any combination of: *FAE1* to produce a low erucic acid phenotype; *TT8* to produce a lower fiber phenotype; and [ ] to produce a lower glucosinolate phenotype. The first two PTMOAs are described in 22-069-01rsr. The third PTMOA is described in 22-241-01rsr. This RSR requests the evaluation of the combination of those previously submitted PTMOAs in pennycress.

Additionally, these traits were evaluated in the AIR process. In response to each of three previously submitted AIR letters<sup>1</sup>, BRS deregulated pennycress lines harboring disruptions in *FAE1*, *TT8*, [ ], and *AOP* genes, concluding that: "...your gene-edited pennycress lines are not themselves plant pests....Additionally, pennycress is not listed as a Federal noxious weed pursuant to 7 CFR part 360 and USDA has no reason to believe that the intended phenotypes of your pennycress lines would increase the weediness of pennycress." *AOP2* is one of several genes that, when disrupted, reduce glucosinolate levels. This RSR requests evaluation of pennycress with mutations in one other gene that results in the same low glucosinolate phenotype, in addition to the same mutations in *FAE1*, *TT8*, and [ ] that lead to the same phenotypes as previously reviewed.

### 1. Information about Requestor

**First Name:** Traci

**Last Name:** Hagedorn

**Position:** Consultant

**Organization Name (if applicable):** Hjelle Advisors, LLC on behalf of CoverCress Inc.

#### Contact information (choose one or both)

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<sup>1</sup> Cover Cress, Inc. Loss of *FAE1*, *TT8*, *AOP2* – USDA response January 29, 2020; CoverCress, Inc. Loss of *FAE1*, *TT8*, [ ] – USDA response May 7, 2020; CoverCress, Inc. Loss of *TT8*, *AOP2* – USDA response August 31, 2020

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## 2. Does the request contain Confidential Business Information (CBI)?

Yes, this RSR request contains CBI.

This RSR for gene-edited pennycress contains confidential business information that could harm CoverCress, Inc. if publicly disclosed prior to publication of patent applications or other disclosures with that information. The specific information that could cause competitive harm are the names and sequences of the low-glucosinolate target genes, as well as the specific modifications and the mechanism of action. Premature disclosure of that information could allow competitors to develop competitive products much more quickly than if the information was kept as CBI and/or trade secret.

CoverCress, Inc. customarily and actually treats commercial and financial information as private and provides these information to the government under an assurance of privacy. We consider the information as confidential within the meaning of 5 U.S.C. §552(b)(4), the Freedom of Information Act's Exemption 4.

## 3. Description of the comparator plant

**Scientific name (genus, species):** *Thlaspi arvense*

**Common Name:** pennycress; field pennycress; CoverCress™

**Subspecies / Cultivar / Breeding Line:**

CoverCress, Inc. has developed through selection and breeding several elite germplasm lines that have optimized agronomic characteristics. This improved germplasm is referred to as CoverCress™ and lines to date include B3, B28, B48, 2032, 183002-B-14, 183002-B-6, 183011-B-5, 183056-B-29, 182002-B-B-31, 183002-B-6, 183002-B-9, 183006-B-13, 183007-B-26, 183007-B-3, 183036-B-10, 183059-B-24, 183059-B-4 and 183059-B-20. It is envisioned that the above-mentioned lines and additional lines developed in the breeding program will undergo late stage editing to produce the lower glucosinolate, low erucic acid, and lower fiber phenotypes within germplasm that is already optimized for commercial production.

## 4. Genotype of the modified plant (genetic material is not inserted into the genome)

**Nature of modification(s)**

Mutations were introduced into pennycress cultivars using a CRISPR/SpCas9 DNA construct designed to target genomic edits to the [ ] *FAE1*, and *TT8* genes. This transgene construct was delivered to the plant using a disarmed *Agrobacterium tumefaciens* strain and a standard floral dip transformation method. When integrated into the plant genome, the expressed *Streptococcus pyogenes* CRISPR-associated protein 9 (SpCas9) endonuclease is guided to four unique, targeted loci (the [ ] *FAE1*, and *TT8* genes). At these locations, the SpCas9 endonuclease catalyzes double-stranded DNA breaks, which are then repaired by the plant's error-prone endogenous non-homologous end joining (NHEJ DNA) repair mechanisms, resulting in heritable mutations at the targeted loci.

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The design pipeline has two steps to ensure the gRNA is specific to the gene and to reduce off-targeting. First, an online gRNA design tool that has an inbuilt pennycress genome is used to identify gRNA target sites and provide a list of putative off-target sites. Furthermore, a blast search is performed with a candidate protospacer sequence against the pennycress genome to avoid the use of a protospacer with potential off-targets.

DsRED fluorescent protein from *Discosoma* was included in the plasmid to confer red fluorescence in plants that successfully take up the plasmid introduced by *A. tumefaciens*. Presence of the construct in T<sub>1</sub> plants was confirmed through visualization of red fluorescent protein under a light system and confirmatory PCR screening of a fragment of the T-DNA. Seed from the progeny T<sub>2</sub> generation was screened for segregants that do not have the transgene as indicated by lack of red fluorescence. Resulting seedlings in the T<sub>2</sub> generation were screened again for negative presence of DsRED and Cas, as well as homozygous edits to [ ] *FAE1*, and *TT8* genes.

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Examples of these edits are listed in the following section.

### Sequence and Comparison of each Modification

[ ]

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The top line of the following sequence comparison is the unmodified sequence of [ ]. One representative modified gene sequence of [ ] is included for comparison. One site in the gene is edited and includes a single base pair insertion, marked in red. The full gene sequences and sequence comparison are included in Appendix 1.

[

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The top line of the following sequence comparison is the unmodified sequence of [ ]. One representative modified gene sequence of [ ] is included for comparison. One site in the gene is edited and includes a single base pair insertion, highlighted in red. The full gene sequences and sequence comparison are included in Appendix 1.

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***FAE1***

The top line of the following sequence comparison is the unmodified sequence of *FAE1*. One representative modified gene sequence of *FAE1* is included for comparison. One site in the gene is edited and includes a single base pair insertion, highlighted in red. The full gene sequences and sequence comparison are included in Appendix 1.

Ta_FAE1	GCCTTCACCGTTTGGCTCTACATCGTAA-CCCGGCCAAACCGGTTACCT
Ta_fael_mut1	GCCTTCACCGTTTGGCTCTACATCGTAA <b>A</b> CCCGGCCAAACCGGTTACCT
	*****

***TT8***

The top line of the following sequence comparison is the unmodified sequence of *TT8*. Two representative modified gene sequences of *TT8* are included for comparison. One site in the gene is edited; one modified sequence includes a single G deletion and one modified sequence includes a single G insertion. The edits are highlighted in red. The full gene sequences and sequence comparison are included in Appendix 1.

Ta_TT8_Wt	GGGAGAATGGATACTACAACGGTGCAATAAG-ACGAGGAAGACAACTCAGCCGGCGGAA
Ta_TT8_Mut2	GGGAGAATGGATACTACAACGGTGCAATAAA <b>-</b> ACGAGGAAGACAACTCAGCCGGCGGAA
Ta_TT8_Mut1	GGGAGAATGGATACTACAACGGTGCAATAAA <b>GG</b> ACGAGGAAGACAACTCAGCCGGCGGAA
	*****

## 5. Description of new traits

Intended trait #1: low total glucosinolate seeds

Intended phenotype: low accumulation of total glucosinolates in seeds; reduction of total glucosinolate levels to below 80 µmol/g of total seed weight

Pennycress contains high levels of oil (~25-35%) that makes it a desirable ultra-low carbon fuel feedstock (Moser et al. 2009; Altendorf et al. 2019; McGinn et al. 2019). In addition to this primary value for fuel, the seed could provide an energy source for animal feeds such as chicken feed. Field pennycress or CoverCress also contains 80-110µmol/g glucosinolates derived through the aliphatic glucosinolate pathway (Chopra et al. 2018b; Chopra et al. 2020; Sedbrook et al. 2014). Glucosinolates are biologically active compounds found in the Brassicaceae family of plants, including broccoli, cabbage, cauliflower, rapeseed, mustard, and horseradish, and provide defense mechanisms for plants. Over 200 types of glucosinolates are found in brassicas (Prieto et al. 2019); pennycress contains the glucosinolate sinigrin (Chopra et al. 2020). Sinigrin and its metabolite allyl isothiocyanate can reduce palatability of food and feed at lower levels and result in toxic effects at higher levels. Therefore, presence of glucosinolates (which is almost all sinigrin) in field pennycress or CoverCress impacts the ability to include the seed or meal in feed applications above limited inclusion rates (e.g. 4% in broiler diets) (Chopra et al. 2020).

Reducing total glucosinolate levels will make pennycress or CoverCress more susceptible to diseases and environmental conditions than the wildtype. If low-glucosinolate seeds are

scattered during harvest, the likelihood of persistence and germination the following season is reduced compared to the wildtype.

Intended trait #2: low erucic acid seeds

Intended phenotype #2: low accumulation of erucic acid in seeds

Erucic acid in seed oil in homozygous *fae1* mutants is consistently <2% of total fatty acids, compared with >35% of total fatty acids in wild-type pennycress seeds (McGinn et al. 2019; Chopra et al. 2020; Chopra et al. 2019). The fatty acid profile of homozygous *fae1* mutants is comprised predominantly of oleic acid (C18:1), linoleic acid (C18:2) and linolenic acid (C18:3), which have known nutritional and energy value.

Intended trait #3: lower fiber seeds

Intended phenotype #3: yellow seed (as a marker for lower fiber)

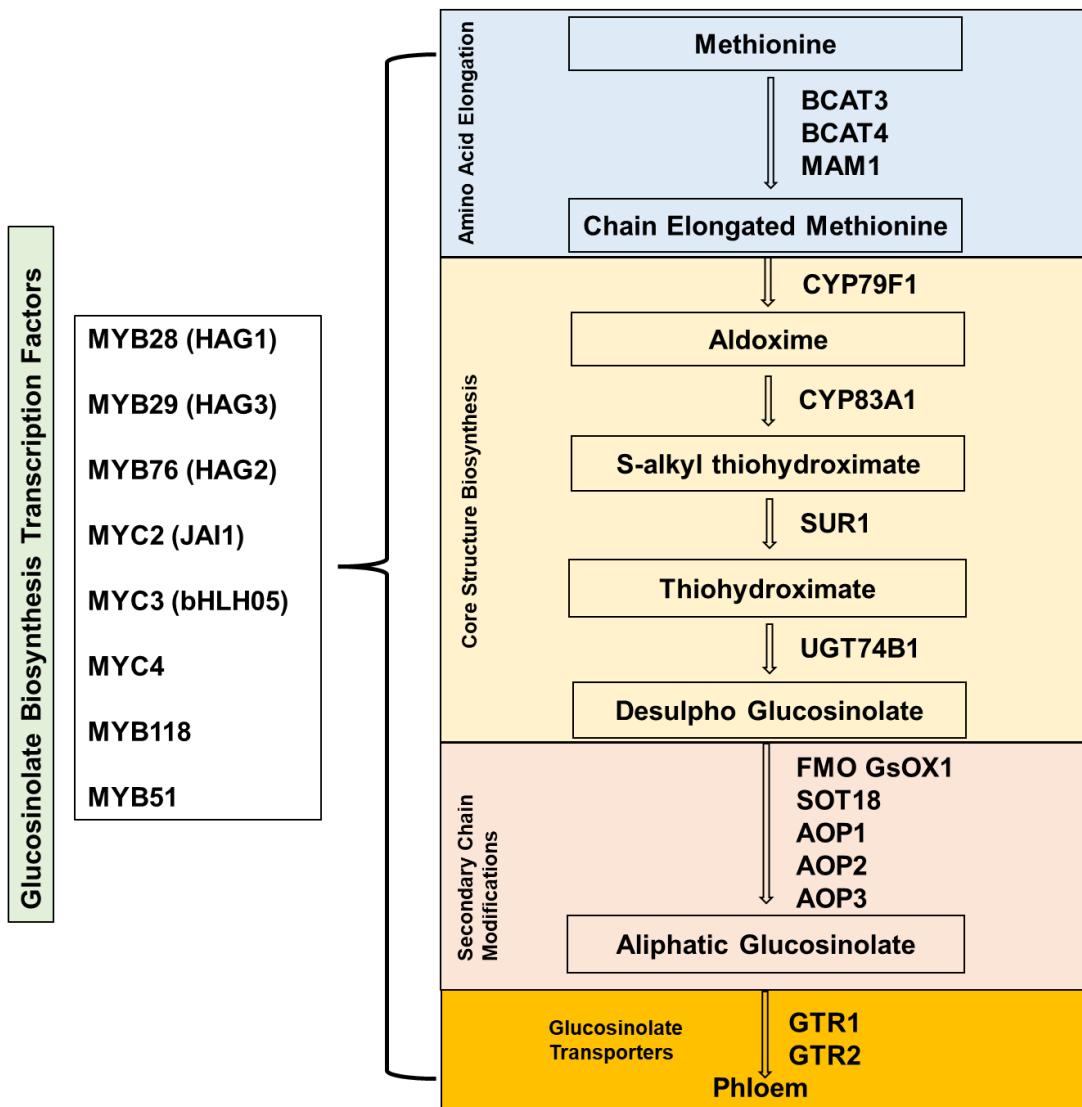
In homozygous *TT8* mutants, the seed coats of pennycress are light yellow colored in contrast to the naturally dark seeds produced by wild-type pennycress, signifying the absence or reduction of condensed tannins in the seed coat. These seeds contain lower levels of undigestible fiber, and thusly, higher metabolizable energy for animal feed.

In a composition study of several light-colored pennycress mutants versus 95 wild type pennycress accessions harvested at various locations across the USA, NIR spectroscopy analysis revealed that the light-colored pennycress contained 10-19.7% Acid Detergent Fiber (ADF) and 13.1-24.1% Neutral Detergent Fiber (NDF), while the dark-colored pennycress contained 20.8-37.9% ADF and 26.3-35.1% NDF (Ulmasov et al. 2020). Additional composition studies consistently show a 25-37% reduction in various fiber components relative to the dark wild type seed (Ulmasov et al. 2020). The lower fiber (yellow seed) phenotype is also associated with reduced seed dormancy, a major factor in the weediness of this plant (Ott et al. 2021; Chopra et al. 2018b).

## 6. Description of each MOA

Low total glucosinolate MOA

Glucosinolates are secondary metabolites important for plant resistance to insects and serve as defense compounds in different tissues of the plant and can help with suppressing weeds. Sønderby et al. (2010) gives an overview of the glucosinolate biosynthesis pathway. Glucosinolate biosynthesis in brassicas requires three steps, outlined in Figure 1: 1) amino acid chain elongation, 2) oxidation and sulfation to generate the core glucosinolate structure and 3) secondary side chain modifications to generate unique forms of glucosinolates. Levels of glucosinolate in the seed are regulated by transporters. There are more than 200 glucosinolates identified and classified in brassicas. Field pennycress or CoverCress contains 80-100 $\mu$ mol/g glucosinolates in the form of sinigrin derived through the aliphatic glucosinolate pathway (Sedbrook et al. 2014; Chopra et al. 2018b; Chopra et al. 2020).



**Figure 1:** Major steps in glucosinolate biosynthesis

The rationale for gene target selection was to identify those genes that result in reduced glucosinolate levels in the seed through a combination of reduced production and mobilization, but do not impact other biosynthetic pathways or cause accumulation of glucosinolates in other parts of the plant. To reach the desired 70-80% reduction in glucosinolate levels will require loss of function mutations in a combination of the 2 gene targets in this RSR request.

These gene targets include the transcription factor [ ] and the transporter [ ]. Each gene and its respective contribution to achieve reduced glucosinolate levels is described below.

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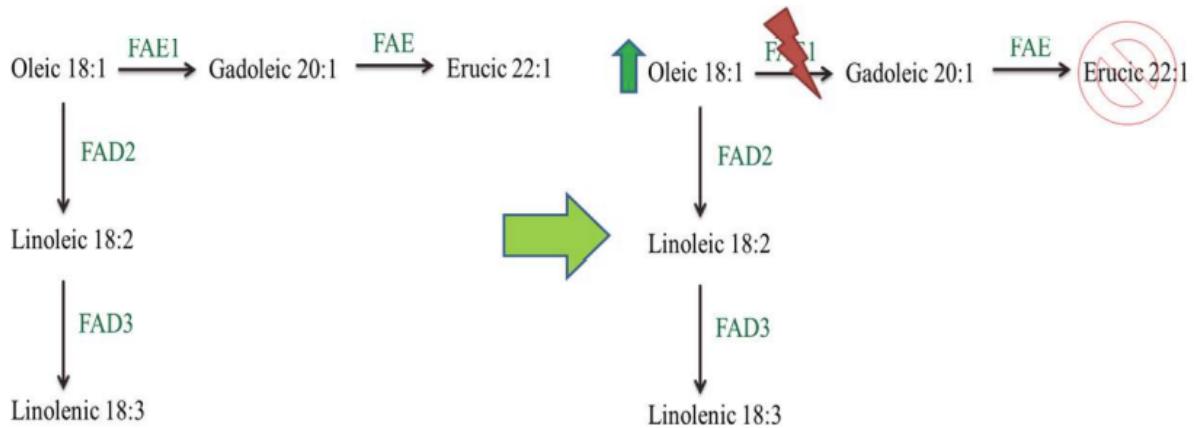
#### Low erucic acid MOA

Field pennycress contains high levels of oil (~25-35%) that makes it a desirable ultra-low carbon fuel feedstock (McGinn et al. 2019; Sedbrook et al. 2014; Moser et al. 2009).

In addition to this primary value for fuel, the seed, meal and/or oil could provide an energy source for animal feeds. The utility of pennycress for this use, however, is limited by that fact that the oil contains >35% erucic acid (Altendorf et al. 2019; McGinn et al. 2019). Erucic acid is a 22-carbon monounsaturated acid that is absorbed, distributed and metabolized like other fatty acids involving primarily metabolism via mitochondrial beta-oxidation and, to a lesser extent, peroxisomal beta-oxidation. Like other longer-chain fatty acids, the rate of mitochondrial beta oxidation is comparatively lower for erucic acid; however, elevated erucic acid levels induce liver peroxisomal oxidation pathways as a mechanism of compensation. Interest in the safety of erucic acid occurred when results of studies in rats associated the dietary intake of high doses of erucic acid with myocardial lipidosis and heart lesions. Oilseed rape conventionally contains similarly high levels of erucic acid. Low erucic acid varieties were identified and marketed as canola, which have been shown to be safe for inclusion in animal feed.

Reduction in erucic acid is achieved through disruption of Fatty Acid Elongation 1 (*FAE1*) (Figure 2), resulting in higher levels of oleic (18:1) (Qiu et al. 2006). It is through this same mechanism that erucic acid levels are lowered in pennycress (McGinn et al. 2019). The edit

to *FAE1* causes a frameshift which results in a premature stop codon and a truncated, non-functional protein.



**Figure 2:** Fatty Acid Pathway to Reduce Erucic Acid Levels

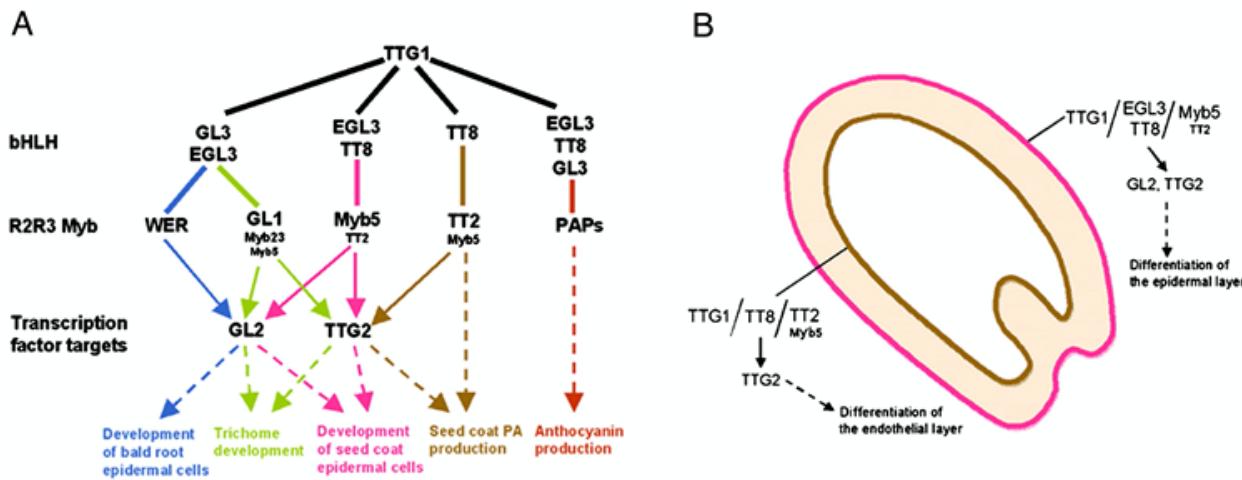
#### Lower fiber MOA

Field Pennycress is high in fiber (level is variable, but Acid Detergent Fraction values of up to 40% mass on a dry weight basis have been observed (Ulmasov et al. 2020)), which can impact digestibility as a feed ingredient.

The production of seed coat fiber was first characterized in the model plant Arabidopsis. Arabidopsis seed coats derive their brown color from the accumulation of proanthocyanidins (PAs), a class of flavonoid chemicals (polymerized flavan-3-ols, or condensed tannins) that protect against a variety of biotic and abiotic stresses and help maintain seed dormancy and viability (Debeaujon et al. 2003). PAs start out as colorless epicatechin compounds until they are transported to the vacuole where they are polymerized and oxidized as the seed desiccates. In Arabidopsis, PAs are only produced in a narrowly defined cell layer in the endothelium of the seed, and TTG1, TT8/bHLH042, and TT2/MYB123 and have been demonstrated as being the three main regulators of PA biosynthesis in seed coat (Baudry et al. 2004; Lepiniec et al. 2006). Gonzalez et al. (2009) described how TTG1 works in a complex with a particular combination of MYB class and bHLH class transcription factors to regulate epidermal development of the seed coat (Figure 3).

Loss-of-function mutants in these genes exhibit the transparent “testa” phenotype as a result of low levels of oxidized PAs in the seed coat (Chopra et al. 2018a). The transparent testa phenotype has been observed in brassicas, including canola, and is characterized by yellow seeds that have more oil because of the resulting thinner seed coat and larger embryo (Abraham and Bhatia 1986). Meal from these brassicas have also been shown to be useful in animal feed because of the relatively lower fiber and higher metabolizable energy (Slominski et al. 1994 and 1999; Simbaya et al. 1995). Similarly, the transparent testa phenotype was observed with loss-of-function mutations in orthologs of these genes in pennycress, resulting

in reduced fiber content (Chopra et al. 2018a). In CoverCress™ the *TT8* gene was edited for loss of function. The edit to *TT8* causes a frameshift which results in a premature stop codon and a truncated, non-functional protein.



**Figure 3:** Models for the regulation of TTG1-dependent pathways in seed coat development. (A) A regulatory network for the positive control of TTG1-dependent epidermal cell fates. Solid lines indicate interactions between members of a complex. Solid arrows indicate direct regulation of *GL2* or *TTG2* targets. Dashed arrows indicate a multi-step differentiation pathway. Colored lines and arrows indicate specific regulator combinations and the pathway controlled. Text size in the case of the MYBs indicates their relative contributions to cell fate regulation. (B) A regulatory model for the differentiation of the seed coat outer and inner layers specified by specific TTG1-dependent transcriptional complexes. Text size in the case of MYB5 and TT2 MYBs indicates their relative contributions to the development of the outer and inner testa layers (Figure from Gonzalez et al. 2009).

In conclusion, CoverCress™ is currently being developed as a new cash cover crop that could be grown between the fall harvest and spring establishment of traditional crops in temperate regions. Edits and/or mutations to [ ] *FAE1*, and *TT8* genes have resulted in the reduced total glucosinolates, low erucic acid, and lower fiber seed traits, respectively. The mechanism of action for the genetic changes leading to each phenotype is well-understood. Based on this information, we request USDA's review of the combination of these previously reviewed PTMOAs and confirmation that the traits, produced through simultaneous or sequential editing of any combination of [ ] *FAE1*, and *TT8* genes in pennycress, is not subject to regulation under Part 340.

## References

Abraham, V. and C. R. Bhatia (1986) Development of Strains with Yellow Seedcoat in Indian Mustard (*Brassica juncea* Czern. & Coss.). *Plant Breeding*, 97; 86–88

Altendorf, K., T. Isbell, D. L. Wyse, and J. A. Anderson (2019) "Significant variation for seed oil content, fatty acid profile, and seed weight in natural populations of field pennycress (*Thlaspi arvense* L.)" *Industrial crops and products* 129: 261-268. doi: 10.1016/j.indcrop.2018.11.054

Baudry, A., M. A. Heim, B. Dubreucq, M. Caboche, B. Weisshaar, L. Lepiniec (2004) *TT2, TT8, and TTG1* synergistically specify the expression of *BANYULS* and proanthocyanidin biosynthesis in *Arabidopsis thaliana*. *Plant J.*, 39, pp. 366-380

Chopra, R., E. B. Johnson, E. Daniels, M. McGinn, K. M. Dorn, M. Esfahanian, N. Folstan, K. Amundson, K. Altendorf, K. Betts, K. Frels, J. A. Anderson, D. L. Wyse, J. C. Sedbrook, M. David Marks (2018a) Translational genomics using *Arabidopsis* as a model enables the characterization of pennycress genes through forward and reverse genetics. *Plant J.* Dec 96(6):1093-1105. doi: 10.1111/tpj.14147. PMID: 30394623

Chopra, R., N. Folstad, J. Lyons, T. Ulmasov, C. Gallaher, L. Sullivan, A. McGovern, R. Mitacek, K. Frels, K. Altendorf, A. Killam, B. Ismail, J. A. Anderson, D. L. Wyse, & M. D. Marks (2018b) The adaptable use of *Brassica* NIRS calibration equations to identify pennycress variants to facilitate the rapid domestication of a new winter oilseed crop. *Industrial crops and products*, 128, 55-61. doi: [10.1016/j.indcrop.2018.10.079](https://doi.org/10.1016/j.indcrop.2018.10.079)

Chopra, R., E. B. Johnson, R. Emenecker, et al. (2020) Identification and stacking of crucial traits required for the domestication of pennycress. *Nat Food* 1, 84–91. <https://doi.org/10.1038/s43016-019-0007-z>

Debeaujon, B. P., R. M. Stubar, D. J. Gingerich, R. D. Vierstra (2003) Proanthocyanidin-accumulating cells in *Arabidopsis* testa: regulation of differentiation and role in seed development. *Plant Cell*, 15. pp. 2514-2531

Gonzalez, A., J. Mendenhall, Y. Huo, A. M. Lloyd (2009) TTG1 complex MYBs, MYB5 and TT2, control outer seed coat differentiation. *Dev. Biol.*, 325, pp. 412-421

Lepiniec, L., I. Debeaujon, J. Routaboul, A. Baudry, L. Pourcel, N. Nesi, M. Caboche (2006) Genetics and biochemistry of seed flavonoids. *Annu. Rev. Plant Biol.*, 57, pp. 405-430

McGinn, M., W. B. Phippen, R. Chopra, S. Bansal, B. A. Jarvis, M. E. Phippen, K. M. Dorn, M. Esfahanian, T. J. Nazarenus, E. B. Cahoon, T. P. Durrett, M. D. Marks, J. C. Sedbrook (2019) Molecular tools enabling pennycress (*Thlaspi arvense*) as a model plant and oilseed cash cover crop. *Plant Biotechnol J.* Apr 17(4):776-788. doi: 10.1111/pbi.13014. Epub 2018 Oct 25. PMID: 30230695; PMCID: PMC6419581

Moser, B. R., S. N. Shah, J. K. Winkler-Moser, S. F. Vaughn, R. L. Evangelista (2009) Composition and physical properties of cress (*Lepidium sativum* L.) and field pennycress (*Thlaspi arvense* L.) oils Industrial Crops and products. Sep 30 (2):199-205. DOI: 10.1016/j.indcrop.2009.03.007

Nour-Eldin, H., T. Andersen, M. Burow *et al.* (2012) NRT/PTR transporters are essential for translocation of glucosinolate defence compounds to seeds. *Nature* 488, 531–534. <https://doi.org/10.1038/nature11285>

Ott, M. A., G. Gardner, K. M. Rai, D. L. Wyse, M. D. Marks, R. Chopra (2021) TRANSPARENT TESTA 2 allele confers major reduction in pennycress (*Thlaspi arvense* L.) seed dormancy, Industrial Crops and Products, Volume 174

Prieto, M. A., C. J. López, J. Simal-Gandara (2019) Glucosinolates: Molecular structure, breakdown, genetic, bioavailability, properties and healthy and adverse effects. *Adv Food Nutr Res.* 2019;90:305-350. doi: 10.1016/bs.afnr.2019.02.008. Epub Mar 25. PMID: 31445598

Qiu, D., C. Morgan, J. Shi, Y. Long, J. Liu, R. Li, et al. (2006) A comparative linkage map of oilseed rape and its use for QTL analysis of seed oil and erucic acid content. *Theor. Appl. Genet.* 114, 67–80. doi: 10.1007/s00122-006-0411-2

Schweizer, F, P. Fernández-Calvo, M. Zander, M. Diez-Díaz, S. Fonseca, G. Glauser, M. G. Lewsey, J. R. Ecker, R. Solano, P. Reymond, (2013) Arabidopsis Basic Helix-Loop-Helix Transcription Factors MYC2, MYC3, and MYC4 Regulate Glucosinolate Biosynthesis, Insect Performance, and Feeding Behavior, *The Plant Cell*, Volume 25, Issue 8, Aug, Pages 3117–3132, <https://doi.org/10.1105/tpc.113.115139>

Sedbrook, J. C., W. B. Phippen, and M. D. Marks (2014) New approaches to facilitate rapid domestication of a wild plant to an oilseed crop: example pennycress (*Thlaspi arvense* L.). *Plant Sci.* 227, 122–132

Simbaya, J, B. A. Slominski, G. Rakow, L. D. Campbell, R. K. Downey, and J. M. Bell (1995) Quality Characteristics of Yellow-Seeded Brassica Seed Meals: Protein, Carbohydrate, and Dietary Fiber Components. *Journal of Agricultural and Food Chemistry* 43 (8): 2062-2066

Slominski, B. A., J. Simbaya, L. D. Campbell, G. Rakow, and W. Guenter (1999) Nutritive value for broilers of meals derived from newly developed varieties of yellow-seeded canola. *Animal Feed Science and Technology* 78:249-262

Slominski, B. A., L. D. Campbell, and W. Guenter (1994) Carbohydrates and dietary fiber components of yellow and brown-seeded canola. *J. Agric. Food Chem.* 42: 704–707

Sønderby I. E., F. Geu-Flores, Halkier, BA (2010) Biosynthesis of glucosinolates – gene discovery and beyond, Trends in Plant Science, Volume 15, Issue 5, 283-290, ISSN 1360-1385, <https://doi.org/10.1016/j.tplants.2010.02.005>

Tan, Z., Z. Xie, L. Dai, Y. Zhang, H. Zhao, S. Tang, L. Wan, X. Yao, L. Guo, D. Hong (2022) Genome- and transcriptome-wide association studies reveal the genetic basis and the breeding history of seed glucosinolate content in *Brassica napus*. Plant Biotechnol J. Jan;20(1):211-225. doi: 10.1111/pbi.13707. Epub 2021 Oct 28. PMID: 34525252; PMCID: PMC8710833

Ulmasov, T., G. Hartnell, J. C. Sedbrook, M. D. Marks, R. Chopra, and M. Esfahanian (2020) Low fiber pennycress meal and methods of making (U.S. Patent No. 10,709,151). U.S. Patent and Trademark Office

**Appendix 1 – Full Sequences and Sequence Comparisons**

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*TaFAE1* Wild Type CDS;

ATGACGTCCGTTAACGTTAAGCTCCTTACCATTACGTACCAACTTTCAACCTTGCTTCTCCCG  
TTAGCGGCGATCGTTGCCGGAAAAGCCTCTCGGCTTACCAACAAACGATCTTCACCACTTCTACTATTCC  
TCTCCAACACAACCTAATAACCATACTCTACTCTTGCCTCACCGTTTGGTT **TGGCTCTACATCGT**  
**AACC** CGGCCAAACCGTTACCTCGTTGACCATTCTGCTACCTTCCACCATCGCATTTAGAAGCAGTA  
TCTCTAAGGTATGGATATCTTCTATCAAGTAAGATTAGCCGATCCTTACGGAACGCGGCAAGCGATGA  
TTCGTCTGGCTTGATTCTTGAGGAAGATTAGGAGCGGTCTGGTAGGCGATGAAACCCACGGCCC

CGAGGGACTGCTTCAGGTCCCTCACGGAAGACTTTGCCGCGCGTGAAGAAACAGAGCAAGTGA  
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*TaFAE1* Mutant1 CDS;

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 AA(A)CCC CGGCCAAACGGTTACCTCGTTGACCATTCTGCTACCTCCACCATCGCATCTAGAACGCA  
 GTATCTCTAAGGTATGGATATCTTCTATCAAGTAAGATTAGCCGATCCTTACGGAACCGGGCAAGCGA  
 TGATTGTCCTGGCTGATTCTTGAGGAAGATTAGGAGCGGTCTGGTCTAGGCATGAAACCCACGG  
 CCCCAGGGACTGCTCAGGCCCTCACGGAAGACTTTGCCGCGCGTGAAGAACAGAGCAAG  
 TGATCATCGGTGCGCTCGAAAAACTATTGAGAACACCAAAGTTAACCTAAAGAGATTGGTATACTTGT  
 GGTGAACCTAAGCATGTTAACCGACTCCTCGCTCGCGATGGTTGTTAATACTTCAAGCTCCGA  
 AGAACATCAGAAGCTTAATCTGGAGGAATGGGTTGAGTGCCTCGTTAGGGTGGCT  
 AAGGACTTGTGATGTCCATAAAAACACTTATGCTCTGTGGTAGCACAGAGAACATCACTAACACA  
 TTATGCTGGTATAACAGATCCATGATGGTTGAAATTGCTTGTGGTGGCTGGGGCGCGATT  
 GCTCTCCAACAAGCCGAGGGACCGGAGACGGTCAAGTACAGCTACTTCACACGGTCCGGACGCATAC  
 CGGAGCTGACGACAAGTCTTCCGATGTGCAACAAGAACGAGAGCGAGAGCGTAAAACCGGGGTGT  
 GTTGTCCAAGGACATAACCGGTGTTGCCGGAGAACTGTTAGAAAACATAACAACATTGGTCCG  
 TGGTTCTCCTTTAGCGAGAAATTCTTTCTGTTACCTCATGCCAAGAAACTCTTAAAGACAAGA  
 TCAAACATTACTACGTCCCGATTCAAGCTGCTATCGACCATTGTATTGATGCCGGAGGCAGAGC  
 CGTGATCGATGTGCTACAGAAGAACCTAGGTCTATTGCCGATCGATGTGGAGGCATCTAGGTCAACGTT  
 ACATAGATTGGAACACTCGTCTAGCTCAATTGGTATGAATTGGCGTACATAGAGGCAAAGGAAG  
 GATGAAGAGAGGGAAACAAAGTTGGCAGATTGCTTAGGGTCAGGGTTAAGTGTAAATAGTGCCTGGTT  
 GGGTGGCTCTACGCAATGTCAAGGCTCGACAAATAGTCTGGAACATTGATTGATAGATATCCAG  
 ATGCAATTGATTCTGATTGGTAAGTCAGAGACTCGTCCAAAACGGTCGGTCTAA

TT8 Wild Type, unmodified sequence

>Ta\_TT8\_Wt

ATGGATGAATCAAGTATTTACGGCAGAGAAAGTGATCGGAGCTGAGAAAAGAGAGCTCAAGGGCT  
GCTTAAGGCCGGCGGTGCAATCTGTGGAGTGGACTTATAGTCTCTTCTGGCAACTTGTCCCTCAACAAAGg  
ttcttttttttaataaaattcatcgatctcacaataaaaaccctaaatttatcatttattatatgttaactacataattatcag  
tatttaaccgtccatgtgcattttggcacttctgtcaattgagttactgaagtaattatatgttaaatgaatttagGGTTTGCTGTG  
GGAGAATGGATACTACAACGGTGCATAAAAGACGAGGAAGACAACTCAGCCGGCGGAAGTGACGGCG  
GAAGAGGCTGCGTTAGAGAGGAGTCAGCAGCTAAGGGAACTTACGAGGCCCTTGGCCGGAGAGTC  
CTCATCGGAAGCTAGGGCATGCACGGCATTATGCCGGAGGATCTGACGGAGACTGAATGGTTTATCT  
AATGTGTGTCCTTCTCTTCCCTCCTCCGGtacccactctctctctctctctctctctctctctctct  
tctctctctttgtctatactgaagttctaatttatctttatcatctctactgaagacaaaaatagtattgtgttaatgcata  
cgaatattgtgaagcattaaaaacaaactgaggagggtgagttactgaaagaagaatgtattggagttgatgaaacgtacactccat  
tttagtgaacataattggaccgttgagattcttattttgtctgattctaaagttagaagcataaaatagataataatgcata  
acaaattgttagttatggatagttatgctttctatgagagaaaaaaaatataatataatgtgaagtaataatttgta  
gGATGCCAGGAAAGGCGTATCGAGGAGGAAACACGTATGGCTATGTGGTGCATAATGAGGTTGACAGT  
AAAATCTTCTAGGGCTATTCTCGCAAAGGtctatttcatttaccactactctatgcatctacttctcacctattt  
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ATCCAGgttaaacgttcttattgattaattctaatttgatgtaatatttacatttacatgtttgaaaattgtttgtataaaaaaa  
aaagCAGACAGTGGTTGCATCCCAGCTTGTGGCGTTGGAACTAGGCACAAACGAACAAAGgtacggc  
gtagttatctttatataatgcataaccaaattgttagaaaaaaaggttagaagagaaatagatcatgcttaagtttatcgttaattaaa  
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ataaaaatattgtttggagatagtcataattctcacaataaaaaataacaaaggatgattaagggaaggagttgatcatgt  
gtttgtctgtgtgtgaagGTAAAAGAAGATATAGCGTTGAGCTATAAAGAGTTTCCATAACCACC  
CCAAGTCAAACCCAAAAGCTGCTTCTGAACACTCCATCACGAAGAGCACGAAGAACGAAAC  
AAGAAGAAGAAGAAGAAGAAGAAGTAGAAGAAGAAATGACAATGTCAGAGGAGATAAGGCTTGGCTC  
TCCTGATGATGATGACGTCTCCAATCAAAACCTACTCTGATTCCATGTAGAATCAACCCACACTTAG  
gtatacacttatacattaaattgttaacgatattcattacacgtatcttattttgttaacaagaaatttttttttttttttttttttt  
tatgtctaaagaaaatctataaaattttatgaatagACACACACATGGACATGATGAATCTAATGGAGGAGGGTGG  
AACTATTCTCAGACAGTATCAACACTTCTTATGTCAACACCCACGAGTCTTTTCAGATTCAAGTTCCACA  
TCTTCTTACATCCAATCATCATTGCCACATGGAAGGCTGATAATTAAAGAGCATCAGCGAGTGGAAA  
CTAAATCGACGTCGTCGCAATGGATGCTAAACACATAATCTGAGAGTCCCTTACTCCACGACCA  
CACTAAAGAAAAGAGGCTGCTCGAGAAAGAGCTTAATCACGTGGTGGCAGAGCGCCGAGGAGAG  
AAGCTGAATGAGAGATTATAACACTGAGATCATTGGTCCCTTGTGACCAAGATGGATAAAAGTCTCA  
ATTCTGGAGACACCCTAACTACGTAAACCATCTCGAAATAGGGCCAAGAGCTGGAGACTAATCATC  
ACGAACAAAAACATAAGCGGATGCGTAGCTGTAAGGGAAAAACGTGGGAAGAGGTCGTTGAGGTTCC  
ATCATAGAGAGTGTGTTAGAGATGAGATGCGAGTACCGAGATGGTCTATTGCTCGACATCCTC  
AGGTTCTTAAGGAACATGGTATAGAGACTACTGCAGTCATACCGCGGTGAACGAGCGTGATTGAGG  
CCGAGATAAGGGCTATGGTGAGAGGGAAAGAAACCAAGCATTGCTGAGGTCAAAGAGGCCATCCATCAA  
ACTATATCCAATATTAACATAG

## TT8 Modified Sequence, Representative Sequence #1

&gt;Ta\_TT8\_Mut1

ATGGATGAATCAAGTATTTACGGCAGAGAAAGTGATCGGAGCTGAGAAAAGAGAGCTCAAGGGCT  
GCTTAAGGCCGGCGGTGCAATCTGTGGAGTGGACTTATAGTCTCTTCTGGCAACTTGTCCCTCAACAAAGg  
ttcttttttttaataaaattcatcgatctcacaataaaaaccctaaatttatcatttattatatgttaactacataattcag  
tatttaaccgtccatgtcattttggcacttctgtcaattgagttactgaagtaattatatgttaaatgaatttagGGTTTGCTGTG  
GGAGAATGGATACTACAACGGTGCAATAAAG(G)ACGAGGAAGACAACTCAGCCGGCGGAAGTGACGG  
CGGAAGAGGCTGCGTTAGAGAGGAGTCAGCAGCTAAGGGAACTTACGAGGCCCTTGGCCGGAGA  
GTCCTCATCGGAAGCTAGGGCATGCACGGCATTATGCCGGAGGATCTGACGGAGACTGAATGGTTTA  
TCTAATGTGTCTCTTCTCTTCCCTCCCTCCGGtacccaactctctctctctctctctctctctctctctct  
tctctctctctctttgtctatactgaagttctaatttatcttttcatctctactgaagacaaaaatagtattgtgttaatgcga  
atcacgaatattgtgaagcattaaaacaaactgaggaggtgagttactgaaagaagaatgtatggagttgatgaaacgtacact  
ccattttgtgaacataattggaccgttgagattttttgtctgatttctaaagttagaagcataaatagataatacataatg  
cataacaaattgttagttatggtagttatgatgatgatgctttctatgagagggaaaaaaaatataatgtgaaagtaataatt  
tttagGATGCCAGGAAAGCGTATCGAGGAGGAAACACGTATGGCTATGTGGTCAAATGAGGTTGA  
CAGTAAAATCTTCTAGGGCTATTCTCGCAAAGgtctattccctttcattaccactactctatgcatctacttcttac  
atttatatatctatcttcaaattaattttctgtcttatttctggatgcttctacatcggtcggttcataatggtagAGTGC  
CAAAATCCAAGttaacgtgtcttattgattaattctatgttagtattttacattttacatgtttgaaaattttgtgata  
aaaaaaaaagCAGACAGTGGTTGCATTCCATGCTTGATGGCGTTGTGGAACTAGGCACAAACGAACAAAGgt  
acggcgtagttatctttatatatgcataacaaatggtaagaaaaaaggttagaagagaaatagatcatgcttaagtttatcagtt  
attaaaaatgtaaaaataagatattatgttcaattatgtatgtccctgttagttaaaaaaaagaataaaaatattaccatttgaag  
tcataatataaaaattgtttggagatagtacataattctcacaattaaaaataacaaaggatgattaaggaaaggatggat  
acatgttggctgtgtgtgaagGTAAGAAGATATAGCGTTGTGAGCTCATAAAGAGTTTCCATAA  
CCACCCCAAGTCAAACCCAAAAGCTGCTTCTGAACACTCCATCAACGAAGAGCACGAAGAACAG  
AGAACACAAGAAGAAGAAGAAGAAGAAGAAGTAGAAGAAGAAATGACAATGTCAGAGGAGATAAGGCTT  
GGCTCTCTGATGATGACGTCTCAATCAAACCTACTCTGATTCCATGTAGAATCAACCCAC  
TTAGgtatacacttatacattaaattgttaacgatattcattacacgtatctttttgttaacaagaattttttattcgccat  
ttctttttatgtctaaagaaaatctataaaattttagatAGACACACATGGACATGATGAATCTAATGGAGGAGG  
GTGGAAACTATTCTCAGACAGTATCAACACTTCTTATGTCAACACCCACGAGTCTTTTCAGATTCTAGTT  
TCCACATCTTACATCCAATCATCATTGCCACATGGAGGCTGATAATTAAAGAGCATCAGCGAGT  
GGAAACTAAATCGACGTCGTCGCAATGGATGCTAAACACATAATCTTGAGAGTCTTCTTACTCCAC  
GACCACACTAAAGAAAAAGAGGCTGCCTCGAGAAGAGCTTAATCAGTGGTGGCAGAGCGCCGAGGA  
GAGAGAAGCTGAATGAGAGATTCTAACACTGAGATCATTGGTCCCTTGTGACCAAGATGGATAAAG  
TCTCAATTCTGGAGACACCCTAACACTACGTAACCATCTCGAAATAGGGCCAAGAGCTGGAGACTAA  
TCATCACGAACAAAAACATAAGCGGATCGTAGCTGTAAGGGAAAAACGTGGGAAGAGGTCGTTGAG  
GTTTCCATCATAGAGAGTGTGTTAGAGATGAGATGCGAGTACCGAGATGGTCTATTGCTCGAC  
ATCCTTCAGGTTCTAAGGAACATGGTATAGAGACTACTGCAGTTCATACCGCGGTGAACGAGCGTGT  
TTCGAGGCCGAGATAAGGGCTATGGTGAGAGGGAAGAAACCAAGCATTGCTGAGGTCAAAGAGCCA  
TCCATCAAACATATCCAATATTAAACTATAG

## TT8 Modified Sequence, Representative Sequence #2

&gt;Ta\_TT8\_Mut2

ATGGATGAATCAAGTATTTACGGCAGAGAAAGTGATCGGAGCTGAGAAAAGAGAGCTCAAGGGCT  
GCTTAAGGCCGGCGGTGCAATCTGTGGAGTGGACTTATAGTCTCTTCTGGCAACTTGTCCCTCAACAAAGg  
ttcttttttttaataaaattcatcgatctcacaataaaaaccctaaatttatcatttattatatgttaactacataattatcag  
tatttaaccgtccatgtcgttattggcacttctgtcaattgagttactgaagtaattatatgttaaatgaatttagGGTTTGCTGTG  
GGAGAATGGATACTACAACGGTGCATAAA(-  
)ACGAGGAAGACAACTCAGCCGGCGGAAGTGACGGCGGAAGAGGGCTGCCTAGAGAGGAGTCAGCAG  
CTAAGGGAACCTTACGAGGCCCTTGCCGGAGAGTCCTCATCGGAAGCTAGGGCATGCACGGCATT  
TCGCCGGAGGATCTGACGGAGACTGAATGGTTTATCTAATGTGTCTCTTCTCTTCCCTCCTTC  
CGGtacccaact  
tatcatctcactgaagacaaaatagtatgtgtttaatgcgaatcacaatattggaagcataaaaacaaactgaggaggtt  
gagttactgaaagaagaaatgtattggagttatgtgatgaaacgtacactccatttagtgaacataattggaccgttgagatttttt  
ctgattgattatctaaagttagaagcataaaatagataatacataatgcataacaattgttagttatggtagttatgtaatgctttct  
atgagagaaaaaaaatataatgtgaaagtaataattttagtGATGCCAGGAAAGGCGTATGCGAGGAGG  
AAACACGTATGGCTATGTGGTGCATAATGAGGTTGACAGTAAAATCTTCTAGGGCTATTCTCGCAAAGG  
tctatttccttttcatttaccactactctatgcatacttctcacatttatatatctatctttcaattaatttctgtcttttt  
tggatgctcctctacatcggtcggttcaatggtagAGTGCCAAATCCAGttaacgttgcatttattgattaattctaatttt  
agtaatatttacattttacatgtttgaaaattgtttgtgataaaaaaaaaagCAGACAGTGGTTGCATTCCATGCTTG  
ATGGCGTTGTGGAACCTAGGCACAACGAACAAAGgtacggcgtagttatctttatatatgcataaccaaattgttagaa  
aaggtagaagagaaatagatcatgcttaagtttatcgtttaattttatcgtttaaaaaatgtaaaaataagatattatgttatttt  
ccctgttagttaaaaaaaaagaataaaatattaccatttgaagtataatataaaaattgttttgagatgtacataatttcacaa  
aaaaaaaaataacaaaggatgattaaggagttggatcatgttgcatt  
GCGTTGTTGAGCTCATAAAGAGTTTCCATAACCACCCCAAGTCACACCCAAAAGCTGCTCTTCTGA  
ACACTCCATCACGAAGAGCACGAAGAACAGACGAAGAACAGAACAGAACAGAACAGTAGAA  
GAAGAAATGACAATGTCAGAGGAGATAAGGCTGGCTCTCTGATGATGATGACGTCTCAAATCAAAC  
CTACTCTGATTCATGTAGAATCAACCCACACTTAGgtatacattatacattaaattgttaacgatattcattaca  
cgatctattttttgttaacaagaaatt  
CACACATGGACATGATGAATCTAATGGAGGAGGGTGGAAACTATTCTCAGACAGTATCAACACTCTTA  
TGTCACAACCCACGAGTCTTTTCAGATTCACTTCCACATCTTCTACATCCAATCATCATTGCCACAT  
GGAAGGCTGATAATTAAAGAGCATCAGCGAGTGGAAACTAAATCGACGTCGTCGCAATGGATG  
CTCAAACACATAATCTTGAGAGTTCTTACTCCACGACCACACTAAAGAAAAGAGGCTGCCTCGAGAAG  
AGCTTAATACGTGGTGGCAGAGCGCCGCAGGAGAGAGAGCTGAATGAGAGAGATTCAAACACTGAGA  
TCATTGGTCCCTTGTGACCAAGATGGATAAAGTCTCAATTCTGGAGACACCATCAACTACGTAAACC  
ATCTCGAAATAGGGCCAAGAGCTGGAGACTAATCATCACGAACAAAAACATAAGCGGATGCGTAGCT  
GTAAGGGAAAACGTGGGAAGAGGTCGTTGAGGTTCCATCATAGAGAGTGTATGTTTAGAGATG  
AGATGCGAGTACCGAGATGGTCTATTGCTCGACATCCTCAGGTTCTAAGGAACATGGTATAGAGACT  
ACTGCAGTTCATACCGCGGTGAACGAGCGTGATTGAGGCCGAGATAAGGGCTATGGTGAGAGGGAA  
GAAACCAAGCATTGCTGAGGTCAAAGAGGCCATCCAACTATCCAATATTAAACTATAG

**Sequence Comparisons of the unmodified and modified genes**

[

CBI-deleted

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CBI-deleted copy

[

CBI-deleted

]

[

CBI-deleted

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***FAE1***

The top lines of the following sequence comparison are the unmodified sequence of *FAE1*. One representative modified gene sequence of *FAE1* is included for comparison. One site in the gene is edited and includes a single base pair insertion, highlighted in red.

Ta_FAE1	ATGACGTCGTTAACGTTAACGTTAACCTTACCGATTACGTACATCACCAACTTTAACCTT *****
Ta_fael_mut1	ATGACGTCGTTAACGTTAACGTTAACCTTACCGATTACGTACATCACCAACTTTAACCTT *****
Ta_FAE1	TGCTTCTTCCCGTTAGCGCGATCGTGCCTGGAAAAGCTCTCGGCTTACCAACAAACGAT TGCTTCTTCCCGTTAGCGCGATCGTGCCTGGAAAAGCTCTCGGCTTACCAACAAACGAT *****
Ta_fael_mut1	*****
Ta_FAE1	CTTCACCACCTCTACTATTCCATCTCCAACACAACCTAATAACCATACTCTACTCTTT CTTCACCACCTCTACTATTCCATCTCCAACACAACCTAATAACCATACTCTACTCTTT *****
Ta_fael_mut1	*****
Ta_FAE1	GCCTTCACCGTTTCGGTTGGCTCTACATCGTAA-CCCGGCCAAACCGGTTACCT GCCTTCACCGTTTCGGTTGGCTCTACATCGTAA <b>A</b> CCGGCCAAACCGGTTACCT *****
Ta_fael_mut1	*****
Ta_FAE1	CGTTGACCATTCTGCTACCTCACCATCGATCTTAGAAGCAGTATCTCTAAGGTCA CGTTGACCATTCTGCTACCTCACCATCGATCTTAGAAGCAGTATCTCTAAGGTCA *****
Ta_fael_mut1	*****
Ta_FAE1	GGATATCTCTATCAAGTAAGATTAGCGATCCTTACGGAACCGGAAAGCGATGATT GGATATCTCTATCAAGTAAGATTAGCGATCCTTACGGAACCGGAAAGCGATGATT *****
Ta_fael_mut1	*****
Ta_FAE1	GTCCTGGCTTGATTCTGAGGAAGATTCAAGGAGCGGTCTGGCTAGGCATGAAACCCA GTCCTGGCTTGATTCTGAGGAAGATTCAAGGAGCGGTCTGGCTAGGCATGAAACCCA *****
Ta_fael_mut1	*****
Ta_FAE1	CGGGCCCCGAGGGACTGCTTCAGGTCCCTCCACCGGAAGACTTTGCCGCGCGGTGAAGA CGGGCCCCGAGGGACTGCTTCAGGTCCCTCCACCGGAAGACTTTGCCGCGCGGTGAAGA *****
Ta_fael_mut1	*****
Ta_FAE1	AACAGAGCAAGTGTACATCGGTGCGCTCGAAAAACTATTGAGAACACCAAAGTTAACCC AACAGAGCAAGTGTACATCGGTGCGCTCGAAAAACTATTGAGAACACCAAAGTTAACCC *****
Ta_fael_mut1	*****
Ta_FAE1	TAAAGAGAGATTGGTATACTTGTGGTAACCAAGCATGTTAACCGACTCCTCGCTCTC TAAAGAGAGATTGGTATACTTGTGGTAACCAAGCATGTTAACCGACTCCTCGCTCTC *****
Ta_fael_mut1	*****
Ta_FAE1	GGCGATGGTTGTTAATACTTCAAGCTCCGAAGCAACATCAGAAGCTTAATCTGGAGG GGCGATGGTTGTTAATACTTCAAGCTCCGAAGCAACATCAGAAGCTTAATCTGGAGG *****
Ta_fael_mut1	*****
Ta_FAE1	AATGGGTTGTAGTGCCGGCGTTAGCCATTGATCTGGCTAAGGACTTGTGCATGTCCA AATGGGTTGTAGTGCCGGCGTTAGCCATTGATCTGGCTAAGGACTTGTGCATGTCCA *****
Ta_fael_mut1	*****

Ta_FAE1	TAAAAACACTTATGCTCTTGTGGTGAGCACAGAGAACATCACTTACAACATTATGCTGG
Ta_fael_mut1	TAAAAACACTTATGCTCTTGTGGTGAGCACAGAGAACATCACTTACAACATTATGCTGG *****
Ta_FAE1	TGATAACAGATCCATGATGGTTTCGAATTGCTTGTCCGTGTTGGTGGGGCCGCAGATT
Ta_fael_mut1	TGATAACAGATCCATGATGGTTTCGAATTGCTTGTCCGTGTTGGTGGGGCCGCAGATT *****
Ta_FAE1	GCTCTCCAACAAGCCAGGGACCGGAGACGGTCCAAGTACCTACAGCTACTTCACACGGTCG
Ta_fael_mut1	GCTCTCCAACAAGCCAGGGACCGGAGACGGTCCAAGTACCTACAGCTACTTCACACGGTCG *****
Ta_FAE1	GACGCATACCGGAGCTGACGACAAGTCTTCCGATGTGTGCAACAAGAAGACGACGAGAG
Ta_fael_mut1	GACGCATACCGGAGCTGACGACAAGTCTTCCGATGTGTGCAACAAGAAGACGACGAGAG *****
Ta_FAE1	CGGTAAAACCBBBBBGTGTGGTCAAGGACATAACCGGTGTTGCCGGAGAACTGTTCA
Ta_fael_mut1	CGGTAAAACCBBBBBGTGTGGTCAAGGACATAACCGGTGTTGCCGGAGAACTGTTCA *****
Ta_FAE1	GAAAAACATAACAACATTGGGTCCGTTGGTCTTCCTTTAGCGAGAAATTCTTTTT
Ta_fael_mut1	GAAAAACATAACAACATTGGGTCCGTTGGTCTTCCTTTAGCGAGAAATTCTTTTT *****
Ta_FAE1	CGTTACCTTCATGCCAAGAAACTCTTAAAGACAAGATCAAACATTACTACGTCCCGA
Ta_fael_mut1	CGTTACCTTCATGCCAAGAAACTCTTAAAGACAAGATCAAACATTACTACGTCCCGA *****
Ta_FAE1	TTTCAAGCTGCTATGACCATTGGTATTGATGCCGGAGGCAGAGCCGTGATCGATGT
Ta_fael_mut1	TTTCAAGCTGCTATGACCATTGGTATTGATGCCGGAGGCAGAGCCGTGATCGATGT *****
Ta_FAE1	GCTACAGAAGAAACTTAGGTCTATTGCCGATCGATGTGGAGGCATCTAGGTCAACGTTACA
Ta_fael_mut1	GCTACAGAAGAAACTTAGGTCTATTGCCGATCGATGTGGAGGCATCTAGGTCAACGTTACA *****
Ta_FAE1	TAGATTGGGAACACTTCGTCTAGCTAATTGGTATGAATTGGCGTACATAGAGGCAA
Ta_fael_mut1	TAGATTGGGAACACTTCGTCTAGCTAATTGGTATGAATTGGCGTACATAGAGGCAA *****
Ta_FAE1	AGGAAGGATGAAGAGAGGGAAACAAAGTTGGCAGATTGCTTGGGTCAAGGGTTAACGT
Ta_fael_mut1	AGGAAGGATGAAGAGAGGGAAACAAAGTTGGCAGATTGCTTGGGTCAAGGGTTAACGT *****
Ta_FAE1	TAATAGTGCCTTGGGTGGCTCTACGCAATGTCAAGGCTTCGACAAATAGTCCTGGGA
Ta_fael_mut1	TAATAGTGCCTTGGGTGGCTCTACGCAATGTCAAGGCTTCGACAAATAGTCCTGGGA *****
Ta_FAE1	ACATTGCATTGATAGATATCCAGATGCAATTGATTGCTGATTGGTAAGTCAGAGACTCG
Ta_fael_mut1	ACATTGCATTGATAGATATCCAGATGCAATTGATTGCTGATTGGTAAGTCAGAGACTCG *****
Ta_FAE1	TGTCCAAAACGGTCGGCCTAA
Ta_fael_mut1	TGTCCAAAACGGTCGGCCTAA *****

**TT8**

The top lines of the following sequence comparison are the unmodified sequence of *TT8*. Two representative modified gene sequences of *TT8* are included for comparison. One site in the gene is edited; one modified sequence includes a single G deletion and one modified sequence includes a single G insertion. The edits are highlighted in red.

Ta_TT8_Wt	ATGGATGAATCAAGTATTTTACGGCAGAGAAAGTGATCGGAGCTGAGAAAAGAGAGCTT
Ta_TT8_Mut2	ATGGATGAATCAAGTATTTTACGGCAGAGAAAGTGATCGGAGCTGAGAAAAGAGAGCTT
Ta_TT8_Mut1	ATGGATGAATCAAGTATTTTACGGCAGAGAAAGTGATCGGAGCTGAGAAAAGAGAGCTT *****
Ta_TT8_Wt	CAAGGGCTGCTTAAGCGGCGGTGCAATCTGGAGTGGACTTATAGTCTCTGGCAA
Ta_TT8_Mut2	CAAGGGCTGCTTAAGCGGCGGTGCAATCTGGAGTGGACTTATAGTCTCTGGCAA
Ta_TT8_Mut1	CAAGGGCTGCTTAAGCGGCGGTGCAATCTGGAGTGGACTTATAGTCTCTGGCAA *****
Ta_TT8_Wt	CTTTGTCCTCAACAAAGGTCTTTTTTTAATAAATTTCATCGATCTCACAATA
Ta_TT8_Mut2	CTTTGTCCTCAACAAAGGTCTTTTTTTAATAAATTTCATCGATCTCACAATA
Ta_TT8_Mut1	CTTTGTCCTCAACAAAGGTCTTTTTTTAATAAATTTCATCGATCTCACAATA *****
Ta_TT8_Wt	AAAACCCAAATTATATCATTATTATTATGTTAACACATAATTATCAGTATT
Ta_TT8_Mut2	AAAACCCAAATTATATCATTATTATTATGTTAACACATAATTATCAGTATT
Ta_TT8_Mut1	AAAACCCAAATTATATCATTATTATTATGTTAACACATAATTATCAGTATT *****
Ta_TT8_Wt	TAACCGTCCATGTGCTTATTGGTCCATTCTGTCATATTACTTGAGGTTCAGA
Ta_TT8_Mut2	TAACCGTCCATGTGCTTATTGGTCCATTCTGTCATATTACTTGAGGTTCAGA
Ta_TT8_Mut1	TAACCGTCCATGTGCTTATTGGTCCATTCTGTCATATTACTTGAGGTTCAGA *****
Ta_TT8_Wt	CTGCCGAGCACATCTCGTTGTCTCGAACATCTGAGACTTTCGTTATTGGCACTT
Ta_TT8_Mut2	CTGCCGAGCACATCTCGTTGTCTCGAACATCTGAGACTTTCGTTATTGGCACTT
Ta_TT8_Mut1	CTGCCGAGCACATCTCGTTGTCTCGAACATCTGAGACTTTCGTTATTGGCACTT *****
Ta_TT8_Wt	CTGTGTCAATTGAGTTACTGAAGTAATTATGTTAACATGAATTAGGGTTTGCTGT
Ta_TT8_Mut2	CTGTGTCAATTGAGTTACTGAAGTAATTATGTTAACATGAATTAGGGTTTGCTGT
Ta_TT8_Mut1	CTGTGTCAATTGAGTTACTGAAGTAATTATGTTAACATGAATTAGGGTTTGCTGT *****
Ta_TT8_Wt	GGGAGAATGGATACTAACGGTCAATAAG-ACGAGGAAGACAACTCAGCCGGCGAA
Ta_TT8_Mut2	GGGAGAATGGATACTAACGGTCAATAA-ACGAGGAAGACAACTCAGCCGGCGAA
Ta_TT8_Mut1	GGGAGAATGGATACTAACGGTCAATAAAG-ACGAGGAAGACAACTCAGCCGGCGAA *****
Ta_TT8_Wt	GTGACGGCGGAAGAGGCTCGTTAGAGAGGAGTCAGCAGCTAACGGAACTTACGAGGCC
Ta_TT8_Mut2	GTGACGGCGGAAGAGGCTCGTTAGAGAGGAGTCAGCAGCTAACGGAACTTACGAGGCC
Ta_TT8_Mut1	GTGACGGCGGAAGAGGCTCGTTAGAGAGGAGTCAGCAGCTAACGGAACTTACGAGGCC *****
Ta_TT8_Wt	CTTTGGCCGGAGAGTCCTCATCGAACGCTAGGGCATGCACGGCATTATGCCGGAGGAT
Ta_TT8_Mut2	CTTTGGCCGGAGAGTCCTCATCGAACGCTAGGGCATGCACGGCATTATGCCGGAGGAT
Ta_TT8_Mut1	CTTTGGCCGGAGAGTCCTCATCGAACGCTAGGGCATGCACGGCATTATGCCGGAGGAT *****
Ta_TT8_Wt	CTGACGGAGACTGAATGGTTTATCTAATGTGTCTCTTCTCTTCCCTCCTCCTTCC
Ta_TT8_Mut2	CTGACGGAGACTGAATGGTTTATCTAATGTGTCTCTTCTCTTCCCTCCTCCTTCC
Ta_TT8_Mut1	CTGACGGAGACTGAATGGTTTATCTAATGTGTCTCTTCTCTTCCCTCCTCCTTCC *****



Ta_TT8_Wt	TTTACATTTATTTACATGTTGAAAATTGTTTGTGATAAAAAAAAGCAGACAGTGGT
Ta_TT8_Mut2	TTTACATTTATTTACATGTTGAAAATTGTTTGTGATAAAAAAAAGCAGACAGTGGT
Ta_TT8_Mut1	TTTACATTTATTTACATGTTGAAAATTGTTTGTGATAAAAAAAAGCAGACAGTGGT *****
Ta_TT8_Wt	TTGCATTCCCATGCTTGATGGCGTTGTGGAACTAGGCACAAACGAACAAGGTACGGCTAG
Ta_TT8_Mut2	TTGCATTCCCATGCTTGATGGCGTTGTGGAACTAGGCACAAACGAACAAGGTACGGCTAG
Ta_TT8_Mut1	TTGCATTCCCATGCTTGATGGCGTTGTGGAACTAGGCACAAACGAACAAGGTACGGCTAG *****
Ta_TT8_Wt	TTATCTTTTATATATGCATAACCAAATGGTTAAGAAAAAGGTTAGAAGAGAAATAGATC
Ta_TT8_Mut2	TTATCTTTTATATATGCATAACCAAATGGTTAAGAAAAAGGTTAGAAGAGAAATAGATC
Ta_TT8_Mut1	TTATCTTTTATATATGCATAACCAAATGGTTAAGAAAAAGGTTAGAAGAGAAATAGATC *****
Ta_TT8_Wt	ATGCTTAAGTTTATCAGTTAAATTAAAAATGTAAAAATAAGATATTATGTTCATTAATA
Ta_TT8_Mut2	ATGCTTAAGTTTATCAGTTAAATTAAAAATGTAAAAATAAGATATTATGTTCATTAATA
Ta_TT8_Mut1	ATGCTTAAGTTTATCAGTTAAATTAAAAATGTAAAAATAAGATATTATGTTCATTAATA *****
Ta_TT8_Wt	ATGTATAGTCCTGTTAGTTAAAAAAAGAATAAAATATTAAACCATTGAAAGTCATAAT
Ta_TT8_Mut2	ATGTATAGTCCTGTTAGTTAAAAAAAGAATAAAATATTAAACCATTGAAAGTCATAAT
Ta_TT8_Mut1	ATGTATAGTCCTGTTAGTTAAAAAAAGAATAAAATATTAAACCATTGAAAGTCATAAT *****
Ta_TT8_Wt	ATAAAAATATTGTTTGGAGATAGTACATAATTCTCACAAATAAAAAATAACAAAGGG
Ta_TT8_Mut2	ATAAAAATATTGTTTGGAGATAGTACATAATTCTCACAAATAAAAAATAACAAAGGG
Ta_TT8_Mut1	ATAAAAATATTGTTTGGAGATAGTACATAATTCTCACAAATAAAAAATAACAAAGGG *****
Ta_TT8_Wt	ATGATTAAGGGAAGGAGTTGGATACATGTTGTTGCTGTGTGAAGGTAAAAGAAG
Ta_TT8_Mut2	ATGATTAAGGGAAGGAGTTGGATACATGTTGTTGCTGTGTGAAGGTAAAAGAAG
Ta_TT8_Mut1	ATGATTAAGGGAAGGAGTTGGATACATGTTGTTGCTGTGTGAAGGTAAAAGAAG *****
Ta_TT8_Wt	ATATAGCGTTGTTGAGCTCATAAAAGAGTTTTCCATAACCACCCCAAGTCACACCCAA
Ta_TT8_Mut2	ATATAGCGTTGTTGAGCTCATAAAAGAGTTTTCCATAACCACCCCAAGTCACACCCAA
Ta_TT8_Mut1	ATATAGCGTTGTTGAGCTCATAAAAGAGTTTTCCATAACCACCCCAAGTCACACCCAA *****
Ta_TT8_Wt	AAGCTGCTTTCTGAACACTCCATCAACGAAGAGCACGAAGAAGACGAAGAACAGAAG
Ta_TT8_Mut2	AAGCTGCTTTCTGAACACTCCATCAACGAAGAGCACGAAGAAGACGAAGAACAGAAG
Ta_TT8_Mut1	AAGCTGCTTTCTGAACACTCCATCAACGAAGAGCACGAAGAAGACGAAGAACAGAAG *****
Ta_TT8_Wt	AAGAAGAAGAAGAAGAAGTAGAAGAAGAAATGACAATGTCAGAGGAGATAAGGCTTGGCT
Ta_TT8_Mut2	AAGAAGAAGAAGAAGAAGTAGAAGAAGAAATGACAATGTCAGAGGAGATAAGGCTTGGCT
Ta_TT8_Mut1	AAGAAGAAGAAGAAGAAGTAGAAGAAGAAATGACAATGTCAGAGGAGATAAGGCTTGGCT *****
Ta_TT8_Wt	CTCCTGATGATGATGACGTCTCCAATCAAACCTACTCTCTGATTCCATGTAGAACCAA
Ta_TT8_Mut2	CTCCTGATGATGATGACGTCTCCAATCAAACCTACTCTCTGATTCCATGTAGAACCAA
Ta_TT8_Mut1	CTCCTGATGATGATGACGTCTCCAATCAAACCTACTCTCTGATTCCATGTAGAACCAA *****
Ta_TT8_Wt	CCCACACTTAGGTATACACTTACATTAAATTAGTTAACGATATCATTACACGTATCT
Ta_TT8_Mut2	CCCACACTTAGGTATACACTTACATTAAATTAGTTAACGATATCATTACACGTATCT
Ta_TT8_Mut1	CCCACACTTAGGTATACACTTACATTAAATTAGTTAACGATATCATTACACGTATCT *****

Ta_TT8_Wt	ATTTATTTGTTAACAGAAATTAAAAATATGCCATTCTTGTTATGTCTAAAGAA
Ta_TT8_Mut2	ATTTATTTGTTAACAGAAATTAAAAATATGCCATTCTTGTTATGTCTAAAGAA
Ta_TT8_Mut1	ATTTATTTGTTAACAGAAATTAAAAATATGCCATTCTTGTTATGTCTAAAGAA *****
Ta_TT8_Wt	AATCTATAAAATTATGAAATAGACACACACATGGACATGATGAATCTAATGGAGGAGGG
Ta_TT8_Mut2	AATCTATAAAATTATGAAATAGACACACACATGGACATGATGAATCTAATGGAGGAGGG
Ta_TT8_Mut1	AATCTATAAAATTATGAAATAGACACACACATGGACATGATGAATCTAATGGAGGAGGG *****
Ta_TT8_Wt	TGGAAACTATTCTCAGACAGTATCAACACTTCTTATGTCACAACCCACGAGTCTTTTC
Ta_TT8_Mut2	TGGAAACTATTCTCAGACAGTATCAACACTTCTTATGTCACAACCCACGAGTCTTTTC
Ta_TT8_Mut1	TGGAAACTATTCTCAGACAGTATCAACACTTCTTATGTCACAACCCACGAGTCTTTTC *****
Ta_TT8_Wt	AGATTCAGTTCCACATCTTCTTACATCCAATCATCATTGCCACATGGAAGGCTGATAA
Ta_TT8_Mut2	AGATTCAGTTCCACATCTTCTTACATCCAATCATCATTGCCACATGGAAGGCTGATAA
Ta_TT8_Mut1	AGATTCAGTTCCACATCTTCTTACATCCAATCATCATTGCCACATGGAAGGCTGATAA *****
Ta_TT8_Wt	TTTAAAGAGCATCAGCGAGTGGAAACTAAATCGACGTCGTCGCAATGGATGCTCAA
Ta_TT8_Mut2	TTTAAAGAGCATCAGCGAGTGGAAACTAAATCGACGTCGTCGCAATGGATGCTCAA
Ta_TT8_Mut1	TTTAAAGAGCATCAGCGAGTGGAAACTAAATCGACGTCGTCGCAATGGATGCTCAA *****
Ta_TT8_Wt	ACACATAATCTTGAGAGTCCCTTACTCCACGACCACACTAAAGAAAAGAGGCTGCC
Ta_TT8_Mut2	ACACATAATCTTGAGAGTCCCTTACTCCACGACCACACTAAAGAAAAGAGGCTGCC
Ta_TT8_Mut1	ACACATAATCTTGAGAGTCCCTTACTCCACGACCACACTAAAGAAAAGAGGCTGCC *****
Ta_TT8_Wt	AGAAGAGCTTAATCACGTGGCAGAGGCCGCAGGAGAGAGAAGCTGAATGAGAGATT
Ta_TT8_Mut2	AGAAGAGCTTAATCACGTGGCAGAGGCCGCAGGAGAGAGAAGCTGAATGAGAGATT
Ta_TT8_Mut1	AGAAGAGCTTAATCACGTGGCAGAGGCCGCAGGAGAGAGAAGCTGAATGAGAGATT *****
Ta_TT8_Wt	CATAACACTGAGATCATTGGTCCCTTGTGACCAAGATGGATAAGTCTCAATTCTGG
Ta_TT8_Mut2	CATAACACTGAGATCATTGGTCCCTTGTGACCAAGATGGATAAGTCTCAATTCTGG
Ta_TT8_Mut1	CATAACACTGAGATCATTGGTCCCTTGTGACCAAGATGGATAAGTCTCAATTCTGG *****
Ta_TT8_Wt	AGACACCATAACTACGTAACCATCTTCGAAATAGGGTCCAAGAGCTGGAGACTAATCA
Ta_TT8_Mut2	AGACACCATAACTACGTAACCATCTTCGAAATAGGGTCCAAGAGCTGGAGACTAATCA
Ta_TT8_Mut1	AGACACCATAACTACGTAACCATCTTCGAAATAGGGTCCAAGAGCTGGAGACTAATCA *****
Ta_TT8_Wt	TCACGAACAAAAACATAAGCGGATGCGTAGCTGAAGGGAAAACGTGGGAAGAGGTCGT
Ta_TT8_Mut2	TCACGAACAAAAACATAAGCGGATGCGTAGCTGAAGGGAAAACGTGGGAAGAGGTCGT
Ta_TT8_Mut1	TCACGAACAAAAACATAAGCGGATGCGTAGCTGAAGGGAAAACGTGGGAAGAGGTCGT *****
Ta_TT8_Wt	TGAGGTTCCATCATAGAGAGTGATGTTTAGAGATGAGATGCGAGTACCGAGATGG
Ta_TT8_Mut2	TGAGGTTCCATCATAGAGAGTGATGTTTAGAGATGAGATGCGAGTACCGAGATGG
Ta_TT8_Mut1	TGAGGTTCCATCATAGAGAGTGATGTTTAGAGATGAGATGCGAGTACCGAGATGG *****
Ta_TT8_Wt	TCTATTGCTGACATCCTTCAGGTTCTTAAGGAACATGGTATAGAGACTACTGCAGTTCA
Ta_TT8_Mut2	TCTATTGCTGACATCCTTCAGGTTCTTAAGGAACATGGTATAGAGACTACTGCAGTTCA
Ta_TT8_Mut1	TCTATTGCTGACATCCTTCAGGTTCTTAAGGAACATGGTATAGAGACTACTGCAGTTCA *****

Ta_TT8_Wt	TACCGCGGTGAACGAGCGTGATTCGAGGCCGAGATAAGGGCTATGGTGAGAGGGAAGAA
Ta_TT8_Mut2	TACCGCGGTGAACGAGCGTGATTCGAGGCCGAGATAAGGGCTATGGTGAGAGGGAAGAA
Ta_TT8_Mut1	TACCGCGGTGAACGAGCGTGATTCGAGGCCGAGATAAGGGCTATGGTGAGAGGGAAGAA
	*****
Ta_TT8_Wt	ACCAAGCATTGCTGAGGTCAAAAGAGCCATCCATCAAACCTATATCCAATATTAACTATA
Ta_TT8_Mut2	ACCAAGCATTGCTGAGGTCAAAAGAGCCATCCATCAAACCTATATCCAATATTAACTATA
Ta_TT8_Mut1	ACCAAGCATTGCTGAGGTCAAAAGAGCCATCCATCAAACCTATATCCAATATTAACTATA
	*****
Ta_TT8_Wt	G
Ta_TT8_Mut2	G
Ta_TT8_Mut1	G