

Veterinary Health Certificate for Export of Equine Semen from the United States to New Zealand



Veterinary Authority
UNITED STATES DEPARTMENT OF AGRICULTURE

Date of Issue

Certificate Number

1. Consignor:

2. Consignee:

3. Country of Origin:
UNITED STATES

4. Zone of Origin:

5. Country of Destination:
NEW ZEALAND

6. Zone of Destination:

7. Place of Origin:

8. Port of Embarkation

9. Estimated Date of Shipment:

10. Means of Transport:

11. Seal Number:

12. Expected Port of Arrival:

13. Description of Commodity:
EQUINE SEMEN

14. Intended Use:
Artificial insemination:
Other:

15. Total Quantity:

16. Total Number of Packages/Containers:

17. Identification of Commodities:

SEE ATTACHMENT 1

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I,, the undersigned USDA Accredited Veterinarian, certify that the semen described above satisfy(ies) the following requirements:

Eligibility

1. The semen is from equids.
2. The semen is fresh-chilled/frozen and non-genetically modified.

Diagnostic testing, vaccination, and treatment

3. All required laboratory testing was conducted at a laboratory approved to conduct export testing of equine semen exported to New Zealand.
4. Original or copies of laboratory reports, or an endorsed, tabulated summary (see Attachment 2), including test date, type, and results for each donor, are attached to this health certificate.
5. All products and vaccinations administered to donor animals for the purposes of meeting the specific disease requirements of this certificate were administered according to the manufacturer's instruction in the United States or in a country approved to export to New Zealand. Vaccinations were either the final dose of a primary course or the recommended booster to complement the primary.

Semen centre requirements

Name of semen centre: _____
 Address of semen centre: _____

 Centre approval number: _____

6. The semen centre meets the conditions specified in the OIE Code Chapter on general hygiene in semen collection and processing centres.
7. The semen centre was:
 - a. Approved for export by APHIS
 - b. Subject to regular annual inspection by an APHIS veterinarian
 Date of last inspection: _____
 - c. Under the supervision of a USDA accredited semen centre veterinarian.
8. If the donors were transferred from one semen centre approved for export to New Zealand to another centre approved for export to New Zealand without isolation or testing, the following occurred (*delete entire clause as appropriate*):
 - a. Donors were examined by the USDA accredited semen centre veterinarian and showed no clinical sign of disease on the day of entry into the facility.
 - b. Transfer was direct.
 - c. Donors were not in direct or indirect contact with animals of a lower health status.
 - d. The means of transport used was disinfected before use.

Semen donor requirements

9. The semen donors were resident for at least 28 consecutive days at the semen centre prior to collection of the semen for export. During this time, semen donors were not used for natural mating and were isolated from animals not of equivalent health status.
10. On the day of collection, the semen centre veterinarian ensured by clinical examination including that of the external reproductive organs that the donor(s) was free from clinical evidence of infectious diseases transmissible in semen.

Semen collection, processing, storage and transport

11. Semen was collected and processed in accordance with the current recommendations of the OIE Code.

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Veterinary Authority

Date of Issue

Certificate Number

UNITED STATES DEPARTMENT OF AGRICULTURE

12. None of the cryogenic or cooling agents have been previously used in association with any other product of animal origin.
13. Semen is placed in straws, ampoules, pellets, or new or disinfected containers (*delete inapplicable container type and initial*) which are sealed and tamper-evident, and clearly and permanently marked to identify the donor and the date(s) of collection in accordance with the OIE Code.
14. Semen was stored with semen/embryos that were collected and processed in accordance with the Code and with semen from donors of equivalent health status. Containers were held until export in a storage place approved by APHIS.
15. Semen was placed in a transport container that is new or disinfected and free of contamination.
Disinfectant (active chemical) and date of disinfection (*delete and initial if container is new*):

16. The transport container was sealed by either the USDA accredited semen centre veterinarian or an APHIS veterinarian using tamper-evident seals.
Seal number _____
17. If the semen was transferred from one transport container to another (*delete entire clause as appropriate*):
Date of transfer: _____
Reason for transfer: _____
Centre moved from: _____
Centre moved to: _____
Receiving centre veterinarian (name and signature): _____
18. The semen in this consignment originates from (*delete entire clause if semen originates from the United States*) _____ (*insert the name of country of origin if not the United States*), which is approved to export equine semen to New Zealand, and is accompanied by:
- a. a declaration from APHIS that links the semen to the semen being exported and confirms that the semen has been stored as per New Zealand requirements at a facility approved by APHIS; AND EITHER
 - i. a veterinary certificate, certified by the Competent Authority of _____ (*insert name of the country of origin if not the United States*) as meeting New Zealand's requirements; OR
 - ii. a letter from the Competent Authority of _____ (*name of country of origin if not the United States*) stating that the semen meets New Zealand's requirements.

SPECIFIC REQUIREMENTS FOR IDENTIFIED RISK ORGANISMS:

19. **Equine herpesvirus-1 (EHV-1)** [abortigenic and paralytic forms]
- a. Donor animals were kept for the 21 days prior to collection in an establishment where no case of EHV-1 (abortigenic and paralytic forms) was reported during that period; and
 - b. Donor animals showed no clinical sign of EHV-1 infection on the day of collection and during the 21 days prior to collection.
20. **Equine infectious anaemia (EIA)**
- a. Donors showed no clinical sign of EIA on the day of each collection; and
 - i. Donors were kept on premises where no case of EIA has been reported during the 90 days prior to each collection; and
 - ii. Donors were subjected to either an agar gel immunodiffusion (AGID) (Coggins) test as per OIE methodology, or an ELISA test, or as listed in the MPI document: [MPI Approved Diagnostic Tests, Vaccines, Treatments and Post-arrival Testing Laboratories for Animal Import Health Standards \(MPI-STD-TVTL\)](#), not less than 21 days after entry into the collection centre with a negative result.

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Certificate Number

21. Equine viral arteritis (EVA) (*delete as applicable*)

a. Donors were kept in an establishment where no equid has shown any clinical sign of EVA for the 28 days immediately prior to semen collection and showed no clinical sign of EVA on the day of semen collection; AND

i. Were subjected between 6 and 9 months of age to a virus neutralization (VN) or reverse transcriptase PCR (EDTA whole blood) test for EVA as per OIE methodology, or as listed in MPI-STD-TVTL, with either (*delete as applicable*)

a) A negative result, OR

b) A positive result, followed at least 14 days later by a second test that showed a stable or decreasing titre; AND

were subsequently vaccinated against EVA and regularly vaccinated according to the recommendations of the manufacturer;

Vaccine name: _____ Vaccination date: _____

OR

ii. Were isolated and not earlier than 7 days after commencing isolation, were subjected to a virus neutralization (VN) or reverse transcriptase PCR (EDTA whole blood) test for EVA as per OIE methodology, or as prescribed in MPI-STD-TVTL, on a blood sample with negative results, vaccinated for EVA, kept for 21 days following vaccination separated from other equids and regularly revaccinated according to the recommendations of the manufacturer;

Vaccine name: _____ Vaccination date: _____

OR

iii. Were subjected to a virus neutralization (VN) or reverse transcriptase PCR (EDTA whole blood) test for EVA as per OIE methodology, or as prescribed in MPI-STD-TVTL, on a blood sample with negative results within 14 days prior to semen collection, and had been separated from other equids not of equivalent health status for 14 days prior to blood sampling until the end of semen collection;

OR

iv. Have been subjected to a virus neutralization (VN) or reverse transcriptase PCR (EDTA whole blood) test for EVA as per OIE methodology, or as prescribed in MPI-STD-TVTL, on a blood sample with positive results, and then EITHER

a) Were subsequently test mated to two mares within 6 months prior to semen collection, which were subjected to two virus neutralization (VN) or reverse transcriptase PCR (EDTA whole blood) tests for EVA as per OIE methodology, or as prescribed in MPI-STD-TVTL, with negative results on blood samples collected at the time of test mating and again 28 days after test mating; OR

b) Were subjected to an agent identification test for EVA, or as prescribed in MPI-STD-TVTL, with negative results, carried out on semen collected within 6 months prior to collection of the semen to be exported; OR

c) Were subjected to an agent identification test for EVA, or as prescribed in MPI-STD-TVTL, with negative results, carried out on semen collected within six months after the blood sample was collected then immediately vaccinated, and revaccinated regularly;

Vaccine name: _____ Vaccination date: _____

OR

v. For frozen semen, were subjected with negative results to either

a) A virus neutralization (VN) or reverse transcriptase PCR (EDTA whole blood) tests for EVA as per OIE methodology, or as prescribed in MPI-STD-TVTL, carried out on a blood sample taken not earlier than 14 days and not later than 12 months after the collection of the semen for export; OR

b) An agent identification test for EVA, or as prescribed by MPI-STD-TVTL, carried out on an aliquot of the semen collected immediately prior to processing or on an aliquot of semen collected within 14 to 30 days after the first collection of the semen to be exported.

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Veterinary Authority
UNITED STATES DEPARTMENT OF AGRICULTURE

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Certificate Number

22. Leptospirosis

- a. Antibiotics effective against *Leptospire*s were added to the semen extender/diluent during processing.

Name and concentration of antibiotics: _____

23. *Taylorella* spp. (Contagious equine metritis, CEM) (delete as applicable)

- a. Donors were from the United States which imposes control measures for CEM as described in the Manual, and
 - i. Have had no direct or indirect contact with CEM during the two months prior to collection, and
 - a) Showed no clinical sign of CEM on the day of each collection; and
 - b) Have been subjected to a culture test as per OIE methodology, or as listed in MPI-STD-TVTL, with negative results twice with a 4-7 day interval during the 30 days prior to the collection period; and
 - c) Have been protected against any possibility of contagion since the beginning of the tests; and
 - d) Have not been treated with antibiotics for at least 7 days before commencing the testing and throughout the sample collection period; OR
 - ii. Have previously shown signs of CEM or have been in direct or indirect contact with CEM during the two months prior to collection; AND
 - a) Were treated for CEM; AND
 - b) After treatment, were subjected to a culture test, or as listed in MPI-STD-TVTL, with three swabs (swabbing sites are the prepuce, the urethral sinus and the fossa glandis [including its diverticulum]) taken at 7 day intervals with negative results followed by testing of the first three mares mated or inseminated by the stallion with negative results; AND
 - c) Have been protected against any possibility of contagion since the beginning of the tests.

USDA Accredited Semen Centre Veterinarian:

Name: _____

Address: _____

Date: _____

Signature: _____

APHIS Veterinarian:

Name: _____

Address: _____

Date: _____

Signature: _____

Stamp:

ATTACHMENT 1

IDENTIFICATION OF THE COMMODITIES

Donor name	Donor ID#	Breed	Date of birth	Date of entry into semen collection centre	Date of semen collection	Collection code	Number of straws

Certificate number: _____

ATTACHMENT 2

LABORATORY DIAGNOSTIC TEST SUMMARY TABLE

Donor name	Donor ID#	Equine infectious anaemia (EIA)			Equine viral arteritis (EVA)			Contagious equine metritis (CEM)		
		Test sampling date	Test type	Test result	Test sampling date	Test type	Test result	Test sampling date	Test type	Test result