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## **Michigan State University**

Information Supporting a Regulatory Status Review of Potato Event Kal91.3 Genetically Modified to Decrease Levels of Invertase Providing Protection from Cold-induced Sweetening

Michigan State University is submitting this information to support a Regulatory Status Review by the USDA Animal and Plant Health Inspection Service under 7 CFR Part 340.4

Submitted on behalf of:  
Potato Breeding and Genetics Program  
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United States

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No CBI

Michigan State University does not consider any information contained in this document to be confidential business information or to be a trade secret.

December 6, 2023

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## **Abbreviations and Definitions:**

LB: Left Border

MOA: Mechanism of Action

MSU: Michigan State University

NPTII: Neomycin phosphotransferase II

ONT: Oxford Nanopore Technology

PCR: Polymerase Chain Reaction

RB: Right Border

RSR: Regulatory Status Review

T-DNA: Transfer DNA

VINV or Vinv: potato vacuolar invertase gene

## 1. Confidential Business Information (CBI) Statement

This RSR request does not contain CBI.

## 2. Product Description and Rationale

### 2A. Requester's name and contact information

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### 2B. Description of plant's genus, species

- Order: Solanales
- Family: Solanaceae
- Genus: Solanum
- Species: *Solanum tuberosum* L.

### 2C. Product Description and Rationale

Vacuolar Invertase silencing is a genetic modification that successfully silences invertase which prevents cold-induced sweetening in potatoes. This modification was demonstrated by Bhaskar, Pudota B., et al. (2010). MSU obtained the vacuolar invertase silencing T-DNA within the plasmid pInvBP1, described in the Bhaskar manuscript from the University of Wisconsin, Madison. The T-DNA was designed to down-regulate the potato vacuolar invertase gene (VInv) transcript through RNAi.

## 3. Description of Host Potato Variety

The MSU conventionally bred potato variety, Kalkaska, is high-yielding and has valuable traits including scab resistance and tolerance to blackspot bruises (Douches, D.S. et al. 2009). Kalkaska is a round white chip-processing variety with a medium set of uniform tubers. The tubers have a low level of internal defects. The strength of this variety is its resistance to common scab, combined with high yield potential and chip-processing quality across many environments.

## 4. Description of Modification

### 4A. Circular map of plasmid vector

A circular map of the plasmid pINVBP1 is represented in Figure 1.

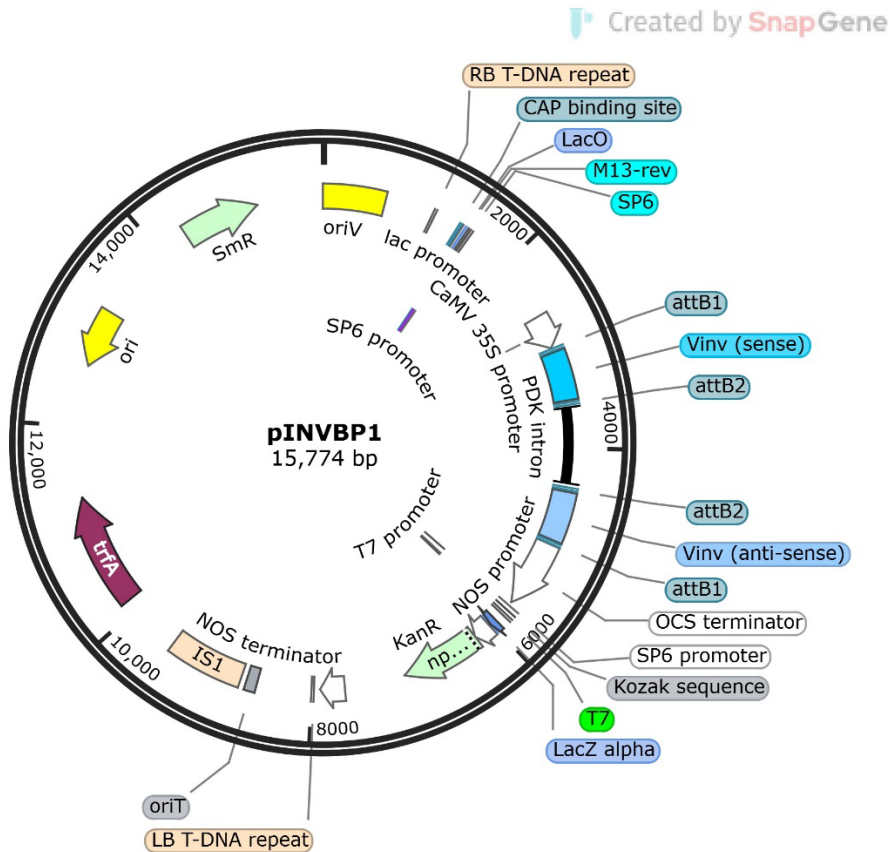


Figure 1. Circular Plasmid Map of pINVBP1.

### 4B. Description of plasmid vector construction

MSU transformed this Kalkaska variety with a plasmid construct developed at UW-Madison called pInvBP1, which contains 508bp of potato VInv cDNA in the plasmid pHELLSGATE 8 using *Agrobacterium*-mediated transfer. The 508-bp cDNA fragment (1,310–1,818 bp; construct InvBP1) was amplified using primers 5'-CACCGAAAGCTTAAGAGGCGGTGATCC-3' (forward) and 5'-CTGCTCCATTCCTGCCTTTGTT-3' (reverse). The amplified PCR products were purified using the QIAquick PCR Purification Kit (Qiagen), gel verified, and cloned into pENTR/D directional TOPO cloning vector (Invitrogen). The VInv cDNA fragments were then transferred into the pHellsGate8 vector using the LR Clonase recombination method (Helliwell et al., 2002). Sequences in the recombinant pHellsGate8-VInv plasmids were confirmed by restriction digestion (*Xba*I and *Xba*I) and sequencing of inserts to ensure that the VInv sequences recombined in sense and antisense orientations. Information on the genetic elements in the plasmid pInvBP1 can be found in Table 1. The Simplot Z6 event, obtained non-regulated status

(Docket No. APHIS–2020–0048), uses a vacuolar invertase silencing cassette containing a tuber-specific promoter called the granule-bound starch synthase (pGbss) gene promoter. Our vacuolar invertase silencing cassette utilizes the commonly used constitutive 35S CaMV promoter.

#### 4C. Sequence of the T-DNA insert

Due to the size of the insert the data has been placed in Section 8. Supplementary Information. Figure 2 on page 13.

#### 4D. Annotation of the T-DNA Inserted Genetic Material

An annotation of the T-DNA of pINVBP1 is described in Table 1.

**Table 1. Genetic elements of the DNA Insert of pInvBP1. T-DNA Right Border (RB) site to Left Border (LB) highlighted in Blue. pINVBP1 plasmid backbone in grey.**

Acronym	Location in pInvBP1	Size (bp)	Direction	Function	Origin
oriV	1..636	636	==		pHELLSGATE Sequence ID: AJ311874.1
Intervening sequence	637..1123	486		Used for cloning	pHELLSGATE
RB T-DNA repeat	1124..1148	25	==	T-DNA right border	<i>Agrobacterium tumefaciens</i>
Intervening sequence	1149...1402	254		Used for cloning	pHELLSGATE
CAP binding site	1403..1424	23	==	Used for cloning	pHELLSGATE
Intervening sequence	1425..1438	14		Used for cloning	pHELLSGATE
lac promoter	1439..1472	32	=>	Used for cloning	pHELLSGATE
Lac operator	1473..1495	24	==	Used for cloning	pHELLSGATE
Intervening sequence	1496..1500	5		Used for cloning	pHELLSGATE
M13-rev	1501..1521	21	=>	Used for cloning	pHELLSGATE
Intervening sequence	1522..1526	5		Used for cloning	pHELLSGATE
SP6 promoter	1527..1553	27	=>	Used for cloning	pHELLSGATE

<b>Intervening sequence</b>	<b>1554..2612</b>	<b>1059</b>			pHELLSGATE
<b>CaMV 35S promoter</b>	<b>2613..2958</b>	<b>347</b>	=>	35S promoter	Cauliflower mosaic virus
<b>Intervening sequence</b>	<b>2959..2966</b>	<b>7</b>		Used for cloning	pHELLSGATE
<b>attB1</b>	<b>2967..2991</b>	<b>25</b>	=>		bacterial attachment site
<b>Intervening sequence</b>	<b>2992..3007</b>	<b>16</b>		Used for cloning	pHELLSGATE
<b>Vinv (sense)</b>	<b>3008..3515</b>	<b>508</b>	==	Fragment of the acid invertase (sense orientation)	Generates double-stranded RNA that triggers the degradation of invertase transcripts
<b>Intervening sequence</b>	<b>3516..3535</b>	<b>20</b>		Used for cloning	pHELLSGATE
<b>attB2</b>	<b>3536..3560</b>	<b>25</b>	<=		bacterial attachment site
<b>Intervening sequence</b>	<b>3561..3586</b>	<b>26</b>		Used for cloning	pHELLSGATE
<b>PDK intron</b>	<b>3587..4355</b>	<b>769</b>	==		Pyruvate orthophosphate dikinase intron from Flaveria trinervia
<b>Intervening sequence</b>	<b>4356..4397</b>	<b>42</b>		Used for cloning	pHELLSGATE
<b>attB2</b>	<b>4398..4422</b>	<b>25</b>	=>		bacterial attachment site
<b>Intervening sequence</b>	<b>4423..4442</b>	<b>20</b>		Used for cloning	pHELLSGATE
<b>Vinv (anti-sense)</b>	<b>4443..4950</b>	<b>508</b>	==	Fragment of the acid invertase (anti-sense orientation)	Generates double-stranded RNA that triggers the degradation of invertase transcripts
<b>Intervening sequence</b>	<b>4951..4966</b>	<b>16</b>		Used for cloning	pHELLSGATE
<b>attB1</b>	<b>4967..4991</b>	<b>25</b>	<=		bacterial attachment site

Intervening sequence	4992..4999	8		Used for cloning	pHELLSGATE
OCS terminator	5000..5707	708	=>		pHELLSGATE
Intervening sequence	5708..5747	40		Used for cloning	pHELLSGATE
SP6 promoter	5748..5766	19	<=		pHELLSGATE
Intervening sequence	5767..5794	28		Used for cloning	pHELLSGATE
Kozak sequence	5795..5804	10	==		pHELLSGATE
Intervening sequence	5805..5833	29		Used for cloning	pHELLSGATE
T7 promoter	5834..5860	29	<=		pHELLSGATE
M13-fwd	5860..5877	18	<=		pHELLSGATE
Intervening sequence	5878..5947	70		Used for cloning	pHELLSGATE
LacZ alpha	5948..6016	69	=>		pHELLSGATE
Intervening sequence	6017..6033	17		Used for cloning	pHELLSGATE
NOS promoter	6034..6217	184	=>	NOS promoter	<i>Agrobacterium tumefaciens</i>
KanR	6218..7039	822	=>	Kanamycin resistance gene	<i>Escherichia coli</i>
Intervening sequence	7040..7663	624			Cloning vector pHELLSGATE
NOS terminator	7664..7916	253	=>	NOS terminator	<i>Agrobacterium tumefaciens</i>
Intervening sequence	7917..7978	62		Used for cloning	pHELLSGATE
LB T-DNA repeat	7979..8003	25	==	T-DNA left border	<i>Agrobacterium tumefaciens</i>
Intervening sequence	8004-8557	554			pHELLSGATE
oriT	8558..8667	110	==		pHELLSGATE
Intervening sequence	8668..8726	59			pHELLSGATE
IS1	8727..9494	768	==		pHELLSGATE
Intervening sequence	9495..10,118	624			pHELLSGATE
trfA	10,119..11,267	1149	=>		pHELLSGATE
Intervening sequence	11,268..12,626	1359			pHELLSGATE
ori	12,627..13,215	589	<=		pHELLSGATE
Intervening sequence	13,216..14,311	1095			pHELLSGATE
SmR	14,312..15,100	789	=>		pHELLSGATE



<b>Intervening sequence</b>	<b>15,101..15,774</b>	<b>673</b>			<b>pHELLSGATE</b>
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#### 4E. Description of the Transformation Method

Potato plant events were produced using *Agrobacterium* transformation as part of the at Michigan State University. The *Agrobacterium* strain GV3101 carrying pINVBP1, was used to transform potato internode explants following the method described by Douches et al. (1998). Transformed internode explants were regenerated on medium containing 50 mg/l kanamycin to select for lines containing a T-DNA insert.

#### 4F. Molecular Characterization of Kal91.3

##### T-DNA Insert

A combination of Xdrop™ enrichment technology (Samplix, Denmark) utilizing Nanopore sequencing and followed by Sanger sequencing as well as corroborating studies using droplet digital PCR (ddPCR) showed the presence of two inserts in Kal91.3 following transformation of Kalkaska with pINVBP1. Collectively, the data indicated that the two copies of the T-DNA were inserted at separate loci, which were shown to be on chromosome 1 and chromosome 3. One insert has a truncation of 513bp of the left T-DNA region which includes the entire NOS terminator of the NPTII selection marker gene. The second insert also has a truncation of 381bp of the left T-DNA which includes some of the NOS terminator of the *nptII* selection marker gene.

The Kal91.3 T-DNA sequence data was generated by first utilizing Xdrop™ enrichment technology. The enriched DNA was subjected to debranching followed by library generation with the ONT Ligation Sequencing Kit. The libraries were sequenced using GridION (Oxford Nanopore Inc.) sequencing platform to generate long-read sequencing data. Junction-finding scripts using these sequences and the DM potato reference genome (Potato Genome Sequencing Consortium, 2011; Sharma et al., 2013), specifically PGSC *S. tuberosum* group *Phureja* clone DM1-3 pseudomolecules (v4.04), indicated two insertion sites, corroborating T-DNA copy number by ddPCR. Sanger sequencing was performed on PCR products across the junctions including at least 1 kb of flanking DNA near the left border and right border for both of the inserts. These sequences matched the plasmid T-DNA sequence.

##### Integration Site

There are two T-DNA inserts in Kal91.3. One insert is located in Chromosome 1 at 6820002 and 6820965 with a 963bp chromosomal deletion compared to the PGSC *S. tuberosum* group *Phureja* clone DM1-3 pseudomolecules (v4.04) reference sequence. The insertion does not interrupt or delete any genes. There is a truncation of 513bp of the left T-DNA region which includes the entire NOS terminator of the *nptII* selection marker gene. The second insert is located in chromosome 3 at 55608167 and 55608288 with a 121bp chromosomal deletion. It also does not interrupt or delete any genes when compared with the DM1-3 pseudomolecules (v4.04) reference sequence. There is a truncation of 381bp of the left T-DNA which includes part of the NOS terminator of the NPTII selection marker gene. The NOS terminator defines the end of the

transcription of the *nptII* gene. Since the *nptII* gene is only used in the selection of the event following transformation, its expression does not impact the success of the Kal91.3 event. The NPTII A summary of the Kal91.3 T-DNA inserts is shown in Table 2.

**Table 2. A summary of the Kal91.3 T-DNA inserts, T-DNA deletions, location in the T-DNA and impact.**

T-DNA Insert in Kal91.3	Deletion of T-DNA	Location in pInvBP1	Function Deleted DNA	Impact of Deleted DNA for Event
T-DNA Insert Chr.1	NOS terminator for the <i>nptII</i> gene was deleted	7491-8003 (513bp)	The NOS terminator defines the end of the transcription of the <i>nptII</i> gene for Kanamycin resistance.	No impact. NPTII used as a selectable marker during the initial transformation of the event
T-DNA Insert Chr.3	Part of the NOS terminator for the <i>nptII</i> gene was deleted	7623-8003 (381bp)	The NOS terminator defines the end of the transcription of the <i>nptII</i> gene for Kanamycin resistance.	No impact. NPTII used as a selectable marker during the initial transformation of the event

#### Absence of Backbone

PCR analysis was used to show the absence of large pINVBP1 plasmid backbone sequences in Kal91.3. This was followed by Southern analysis, using 6 PCR labeled probes that span the entire pINVBP1 backbone region, for the detection of small backbone sequences. The results showed that no backbone from the plasmid was inserted in Kal91.3.

## 5 Description of New Trait

### 5A. Intended trait

#### **Invertase silencing:**

The potato contains a vacuolar invertase RNAi silencing cassette that successfully down-regulates the invertase which prevents cold-induced sweetening in potatoes.

#### **NPTII:**

The potato contains a protein that confers resistance to the antibiotic neomycin and serves as a selectable marker for plant transformation.

## 5B. Intended phenotype

### **Invertase silencing:**

Potatoes that have successful down-regulation of invertase can be stored in the cold (4°C) for much longer times than standard 10°C cold storage durations. Cold-stored potatoes without down-regulation are subject to cold sweetening and French fries and potato chips made from these tubers become brown to black when processed (fry method), taste bitter, and may have elevated levels of acrylamide, a possible carcinogen.

### **NPTII:**

Potatoes are resistant to the antibiotic neomycin and the NPTII protein serves as a selectable marker for plant transformation.

## 5C. Description of the Mechanism of Action (MOA)

The following describes the mechanism of action of RNAi Invertase silencing.

Down-regulation of invertase slows the conversion of sucrose into fructose and glucose in the vacuole. Baskar et al.(2010) determined that RNAi silencing of the *VInv* gene does not affect potato development. This MOA has been previously reviewed and given non-regulated status (Docket No. APHIS–2020–0048)

The second gene transferred to the potato genome encodes neomycin phosphotransferase II (NPTII). Expression of this protein in plant cells confers resistance to the antibiotics neomycin and kanamycin and serves as a selectable marker for plant transformation. The antibiotics neomycin and kanamycin, bind to the negatively charged backbone of nucleic acids to disrupt protein synthesis and therefore inhibit bacterial cell growth. Neomycin phosphotransferase II catalyzes the addition of phosphate from ATP to the 3'-hydroxyl group of the 4,6-disubstituted aminoglycosides neomycin and kanamycin. The NPTII-mediated phosphorylation of neomycin/kanamycin introduces a phosphate group on the antibiotic that reduces the binding affinity to nucleic acids due to steric hindrances and unfavorable electrostatic interactions and thereby disrupts the mechanism of action of the antibiotic (Wright and Thompson, 1999).

In the absence of neomycin or kanamycin, the NPTII expressed in the potato transformed event is not expected to exhibit any enzymatic activity. Therefore, NPTII is not expected to have any effect on other potato metabolic pathways. Additionally, the food, feed and environmental safety of NPTII has been well established. Neomycin phosphotransferase II has been used as a selectable marker in many different commercial genetically-engineered (GE) crops {e.g. Genuity® DroughtGard™ corn (MON 87460), YieldGard® Rootworm corn (MON 863), Bollgard® cotton (MON 531), Bollgard®II cotton (MON 15985), Roundup Ready® cotton (MON 1445)}, and therefore has a history of safe use in the environment as well as in food and feed uses. Furthermore, the NPTII protein has been fully characterized, and the NPTII protein expressed in GE crops has been shown not to pose any discernable environmental, food, or feed safety concerns (Fuchs et al., 1993a and 1993b; Nap et al., 1992; Flavell et al, 1992).

## 5D. Metabolism, Physiology, and Development

None of the modifications in Kal91.3 have an impact on the metabolism, physiology, or development of the plant.

## 6 Proposed plant-trait-MOA language for the website

Plant: *Solanum tuberosum* (potato)

Trait: Invertase gene silencing

Phenotype: Decreased cold-induced sweetening in potatoes.

MOA: Down-regulates the amount of invertase, which slows the conversion of sucrose into fructose and glucose in the vacuole.

## 7 Literature Cited

Bhaskar, Pudota B., et al. "Suppression of the vacuolar invertase gene prevents cold-induced sweetening in potato." *Plant Physiology* 154.2 (2010): 939-948. (UW-Madison)

Douches, D.S., Coombs, J., Hammerschmidt, R., Kirk, W.W. and Long, C., 2009. Kalkaska: a round white chip-processing potato variety with common scab resistance. *American Journal of Potato Research*, 86, pp.347-355.

Douches, D.S., A.L. Westedt, K. A. Zarka, and E.J. Grafius. 1998. Transformation of CryV-Bt transgene combined with natural resistance mechanisms for resistance to tuber moth in potato (*Solanum tuberosum* L.) *HortScience* 33(6):1053-1056.

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Fuchs, R.L., Ream, J.E., Hammond, B.G., Naylor, M.W., Leimgruber, R.M. and Berberich, S.A., 1993. Safety assessment of the neomycin phosphotransferase II (NPTII) protein. *Bio/technology*, 11(12), pp.1543-1547.

Nap, J-P., Bijvoet, J. and Stiekema, W.J. (1992) Biosafety of kanamycin-resistant transgenic plants. *Transgenic Research* 1: 239-249.

Potato Genome Sequencing Consortium, 2011. Genome sequence and analysis of the tuber crop potato. *Nature* 475, 189–95. <https://doi.org/10.1038/nature10158>

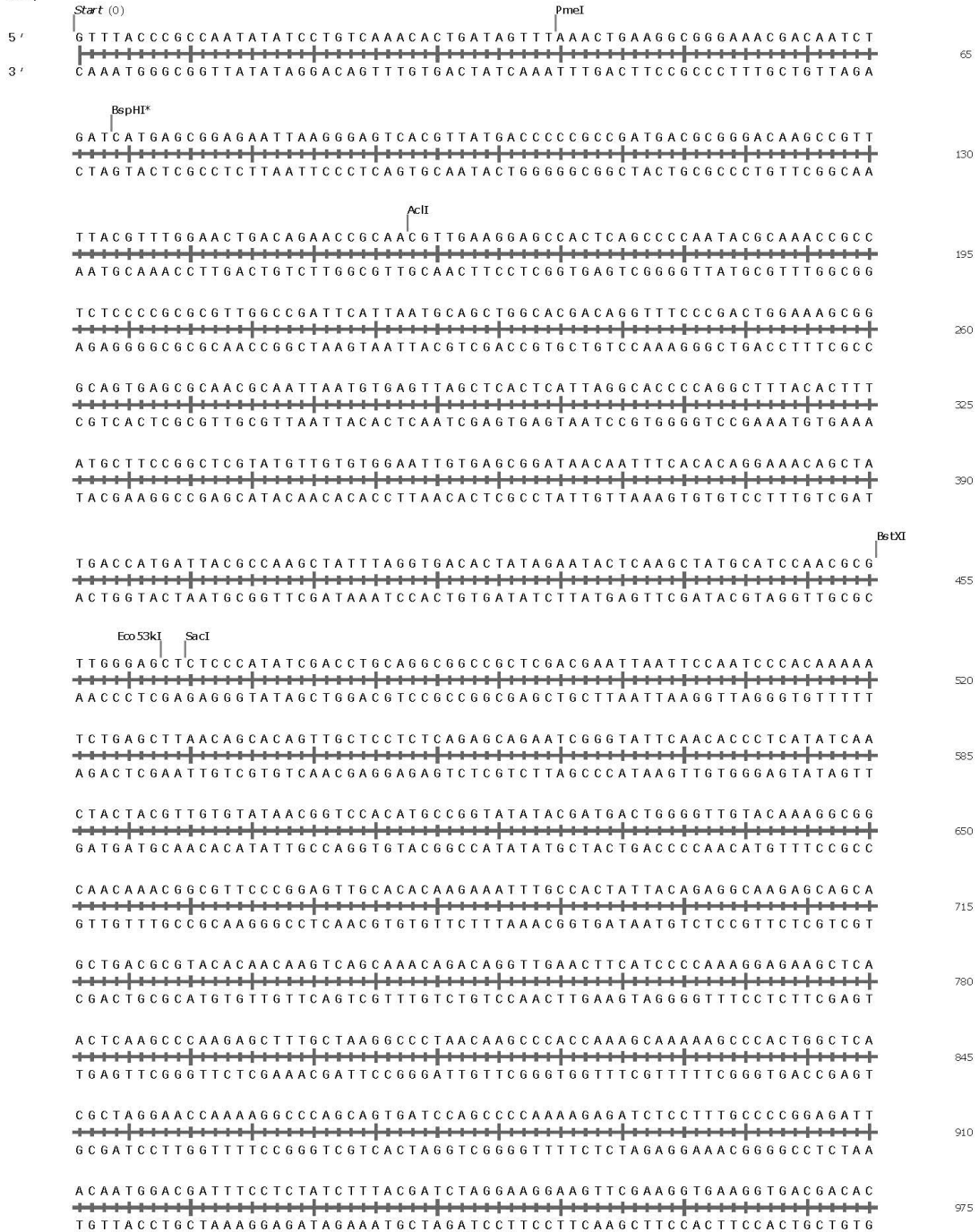
Sharma, S.K., Bolser, D., de Boer, J., Sønderkær, M., Amoros, W., 2013. Construction of reference chromosome-scale pseudomolecules for potato: integrating the potato genome with genetic and physical maps. *Genes Genomes Genet.* 3, 2031–47. <https://doi.org/10.1534/g3.113.007153>

Wright, G.D, and Thompson, P.R. (1999) Aminoglycoside phosphotransferases: proteins, structure, and mechanism. *Front Biosci.* 4:1-3.

## 8. Supplementary Information

**Figure 2. Sequence of the pINVP1 T-DNA insert:**

Sequence: pINVP1 TDNA only (Linear / 6880 bp)  
Enzymes: Unique 6+ Cutters (40 of 678 total)



TATGTTCACTGATAATGAGAAGGTTAGCCTCTTCAATTTTCAGAAAAGAAATGCTGACCCACAGA  
 ATACAAGTGGTGAATACTTCTTCCAATCGGAGAAGTTAAAGTCTTTCTTACGACTGGGTGCT 1040

StuI  
 TGGTTAGAGAGGCCCTACGCAGCAGGTTCTCATCAAGACGATCTACCCGAGTAACAATCTCCAGGAG  
 ACCAATCTCTCCGGATGCGTCTGCCAGAGTAGTTCTGCTAGATGGGCTCATTGTTAGAGGTCCTC 1105

ATCAAAATACCTTCCCAAGAAGGTTAAAGATGCAAGTCAAAAGATTCAAGGACTAATTGCATCAAGAA  
 TAGTTTATGGAAGGGTTCTTCCAATTTCTACGTCAGTTTTCTAAAGTCTGATTAAACGTAGTTCTT 1170

ScaI  
 CACAGAGAAAAGACATATTTCTCAAGATCAGAAGTACTATTCCAGTATGGACGATTCAAGGCTTGC  
 GTGTCCTTTCTGTATAAAAGAGTTCTAGTCTTTCATGATAAAGGTCATACCTGCTAAAGTTCGAAACG 1235

TTCATAAACCAAGGCAAGTAATAGAGATTGGAGTCTCTAAAAAGGTAGTTCTTACTGAATCTAAG  
 AAGTATTTGGTTCCGTTTCTTATCTCTAACCTCAGAGATTTTCCATCAAGGATGACTTAGATTC 1300

GCCATGCAATGGAGTCTAAGATTCAAATCGAGGATCTAACAGAACTCGCCGTGAAGACTGGCGAAC  
 CGGTACGTACCTCAGATTCTAAGTTTAGCTCCTAGATTGCTTGGAGCGGCACCTCTGACCGCTTG 1365

AGTTCATACAGAGTCTTTTACGACTCAATGACAAGAAGAAAATCTTCTGCAACATGGTGGAGCAC  
 TCAAGTATGCTCAGAAAATGCTGAGTTACTGTTCTTTTAAAGCAAGTTGTACCACCTCGTG 1430

AccI  
 GACACTCTGGTCTACTCCAAAAATGTCAAAGATACAGTCTCAGAAGACCAAAGGGCTATTGAGAC  
 CTGTGAGACCAGATGAGGTTTTTACAGTTTCTATGTCAGAGTCTTCTGGTTTTCCCGATAACTCTG 1495

TTTTCAACAAAGGATAATTTTGGGAAACCTCCTCGGATTCCATTGCCAGCTATCTGTCACTTCA  
 AAAAGTTGTTTCTATTAAAGCCCTTTTGGAGGAGCCTAAGGTAACGGGTCGATAGACAGTGAAGT 1560

TCGAAAAGGACAGTAGAAAAGGAAAGGTTGGCTCTACAAATGCCATCATTTGCGATAAAAGGAAAGGCT  
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PshAI  
 ATCATTCAAGATCTCTCTGCGGACAGTGGTCCCAAAGATGGACCCCAACCCACGAGGAGCATCGT  
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BmgBI  
 GGAAAAAGAAGACGTTTCCAAACCAGTCTTCAAAGCAAGTGGATTGATGTGACATCTCCACTGACG  
 CCTTTTCTTCTGCAAAGGTTGGTGCAGAAAGTTTCTGTTACCTAACTACTGATAGAGGTGACTGC 1755

TAAGGGATGACGCACAATCCCACTATCTTTCGCAAGACCTTCTCTATATAAGGAAAGTTTCAATTT  
 ATTCCCTACTGCGTGTAGGGTGTAGGAAAGCGTCTG66AAGGAGATATTTCTTCAAGTAA 1820

CATTTGGAGAGGACACGCTCGAGACAAGTTTGTACAAAAAGCAAGGCTCCGCGGGCCGCCCTTCT  
 GTAAACCTCTCCTGTGCGAGCTCTGTTCAAACATGTTTTTCTGTCGAGGCGCCGGCGGGGAAAG 1885

ACCGAAAGCTTAAAGAGGCGGTGATCCTATTGTTAAGCAAGTCAATCTTCAACCAGGTTCAATTGA  
 TGGCTTTCGAATTTCTCGCCACTAGGATAACAATTCGTTCAAGTTAGAAAGTTGGTCCAAAGTTAACT 1950

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GCTACTCCATGTTGACTCAGCTGCAGAGTTGGATATAGAAGCCTCATTGAAAGTGGACAAAAGTCG
CGATGAGGTACAACCTGAGTCGACGCTCAACCTATATCTTCGGAGTAAACTTCACCTGTTTCAGC
CGCTCCAGGGAAATAATTGAAGCAGATCATGTAGGTTTCAGCTGCTCTACTAGTGGAGGTGCTGCT
GCGAGGTCCCTTATTAACCTTCGCTAGTACATCCAAAGTCGACGAGATGATCACCTCCACGACGA
AGCAGAGGCCATTTTGGGACCATTGGTGTCTGTTGTAATTGCTGATCAAACGCTATCTGAGCTAAC
TCGTCTCCGTAAAACCTGGTAAACCACAGCAACATTAACGACTAGTTTGCATAGACTCGATTG
GCCAGTTTACTTCTACATTTCTAAAGGAGCTGATGGCCGAGCTGAGACTCACTTCTGTGCTGATC
CGGTCAAATGAAAGATGTAAGATTCTCTGACTACCGGCTCGACTCTGAGTGAAGACACGACTAG
AAACAGATCTCTCAGAGGCTCCGGGAGTTGCTAAACAAGTTTATGGTAGTTACAGTACCCGTGTTG
TTTGGTCTAGGAGTCTCCGAGGCCCTCAACGATTTGTTCAAATACCATCAAAGTCATGGGCACAAC
GACGGTGAAAAACATTCGATGAGATTATTGGTGGACCACTCAATTGTGGAGAGCTTTGCTCAAGG
CTGCCACTTTTGTAAAGCTACTCTAATAACCACCTGGTGAAGTTAACCTCTCGAAACGAGTTCC
AGGAAGAACAGTGCATAACATCGCGAATTTACCCAACAAGGCAGTGAATGGAGCAGAAAGGGTGGG
TCCTTCTTGTCAAGATTGTAGCGCTTAAATGGGTTGTTTCCGCTCACTTACCTCGCTTCCCACCC
                                EcoRI   Acc65I   KpnI
CGCGCCGACCCAGCTTTCTTGTACAAAAGTGGTCTCGAGGAATTCGGTACCCAGCTTGGTAAAGGA
GC6CGGCTGGGTCGAAAAGAACATGTTTACCAGAGCTCCTTAAGCCATGGGGTCGAACCACTTCT
AATAATTATTTTCTTTTTCTTTTTAGTATAAAAATAGTTAAGTGAATGTTAATTAGTATGATTATA
TTATTAATAAAAAGAAAAAGGAAAAATCATTTTTATCAATCACTACAATTAATCATACTAATAT
ATAATATAGTTGTATAATTGTGAAAAAATAATTTATAAATATATTGTTTACATAAAACAACATAG
TATTATATCAACAATATTAACACTTTTTTATTAATATTTATATAACAATGATTTGTTGATC
TAATGTAAAAAATATGACAAGTGATGTGTAAGACGAAGAAGATAAAAAGTTGAGAGTAAGTATAT
ATTACATTTTTTTATACTGTTCACTACACATTCGCTTCTTCTATTTTTCAACTCTCATTATATA
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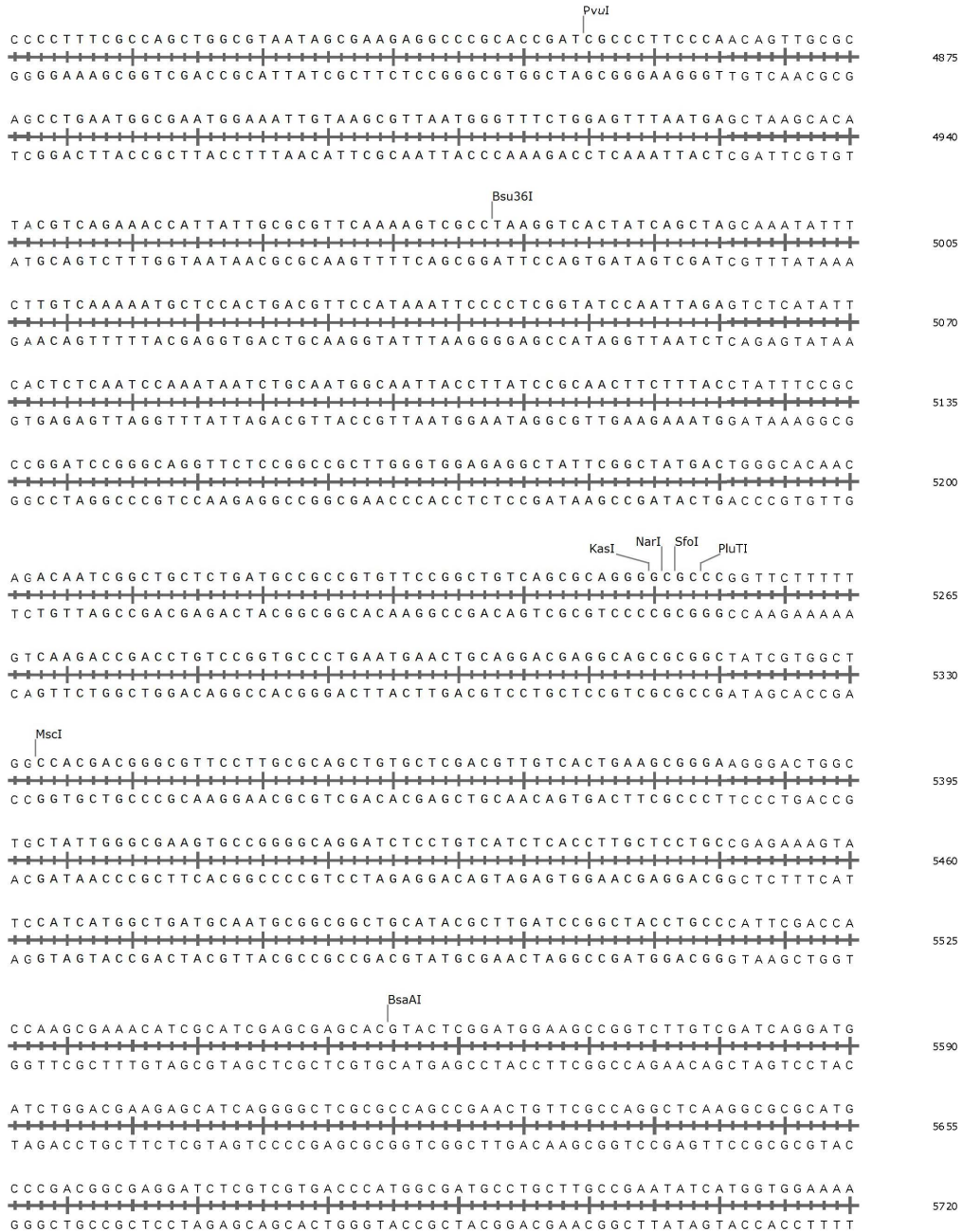
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