United States Department of Agriculture Animal and Plant Health Inspection Service Center for Veterinary Biologics 1800 Dayton Avenue Ames, IA 50010

1. Reagent Name: Clostridium perfringens type C (beta) toxin

2. Strain or Source: Not applicable.

3. Lot Number: IRP 513 (04)

4. Fill Date: January 27, 2005

5. Expiration Date: No expiration date has been assigned to this product because *C. perfringens* type C (beta) toxin has demonstrated over time to be very stable if properly stored. The stability of this reagent will be routinely monitored by the Center for Veterinary Biologics.

Precautions: This reagent does not present a hazard to laboratory personnel who work with the toxin provided good fundamental laboratory techniques are followed.

- **6. Intended Use:** IRP 513 (04) serves as the standard toxin when conducting *C. perfringens* type C toxin-neutralization tests in mice.
- 7. Instructions for Use: IRP 513 (04) diluted 1:110 is considered the standard toxin dilution when conducting toxin-neutralization tests in mice as outlined in title 9, *Code of Federal Regulations* (9 CFR), section 113.111, and 9 CFR 113.454. The standard toxin dilution is prepared by adding 1.0 mL of IRP 513 (04) to 9.0 mL of peptone diluent (1.0% peptone, 0.25% sodium chloride, pH 7.2). The toxin is further diluted to 1:110 by adding 1.0 mL of the 1:10 dilution to 10.0 mL of diluent. A volume of 0.5 mL of the toxin diluted 1:110 and 0.5 mL of diluent is equivalent to 10 L_0 doses. A volume of 0.8 mL of toxin diluted 1:110 and 0.2 mL of diluent is equivalent to 10 L_+ doses. *Clostridium perfringens* type C (beta) toxin IRP 513 (04) diluted 1:10 is stable when stored at -70° \pm 10°C.

8. Test of Reagent:

Determination of test dose of toxin - The $10 L_0$ and $10 L_+$ doses were established by injecting mice intravenously with 0.2 mL of a mixture of varying amounts of IRP 513 (04) combined with 10 International units (IU) of *C. perfringens* beta antitoxin. The $10 L_0$ dose for *C. perfringens* type C (beta) toxin neutralization test is the largest amount of toxin which can be mixed with 10 IU of beta antitoxin and not cause death in injected mice within 24 hours. The $10 L_+$ dose for *C. perfringens* type C (beta) toxin neutralization test is the smallest amount of toxin which can be mixed with 10 IU of beta antitoxin and cause death in at least 80% of injected mice within 24 hours.

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Determination of LD_{50} in mice - Female white Swiss mice weighing 16-20 g were injected intravenously with 0.2 mL of IRP 513 (04) diluted in peptone diluent. The toxin was found to contain $10^{4.016}$ mouse lethal dose fifty (LD₅₀) per 0.2 mL.

Determination of toxin type - The toxin type was confirmed by performing toxin neutralization tests in mice. The mice were injected intravenously with mixtures of IRP 513 (04) and *C. perfringens* type A, B, C, or D antitoxin. All of the mice died within 24 hours except those receiving mixtures containing type B or C antitoxin.

Sterility test - The toxin was tested for sterility and found to be free of viable bacteria and fungi according to the procedures outlined in 9 CFR 113.26.

- **9. Container Size, Type, Weight, or Volume:** 4.0-mL glass vials containing 1.3 mL of toxin.
- 10. Storage Conditions: Store at $-70^{\circ} \pm 10^{\circ}$ C.
- **11. CVB Technical Contact:** Bacteriology Section, Center for Veterinary Biologics, (515) 337-6140 or FAX (515) 337-7673.
- **12. Origin and Passage History:** *C. perfringens* type C (beta) culture #4414, used to produce IRP 513 (04), was obtained from Coopers Animal Health, Inc., on July 28, 1975. The number of passages is unknown.
- **13. Method of Preparation:** Culture #4414 was grown in a 14-liter New Brunswick fermentor containing media consisting of N-Z case, proteose peptone, and yeast extract. Actively growing culture was aseptically added to the fermentor and incubated at 35°C for approximately 4 hours. The culture was centrifuged at 10,000 x g for 60 minutes. The supernatant was passed through a Millipore filtration unit containing a 0.2-μm membrane. The filtrate was further processed using a Millipore pellicon cassette system containing a high volume ultrafilter. The concentrated toxin was adjusted to pH 6.6 and passed through a sterile Millipore filtration unit containing a 0.22-mm membrane. Sterile glycerol was added to the product at a final concentration of 15% v/v.

14. Other: None.

Reagent orders and feedback should be sent *including phone number* to the following email address: CVB@aphis.usda.gov

Reagent orders forms (APHIS 2018) are available from: https://www.aphis.usda.gov/library/forms/pdf/APHIS_2018.pdf

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